

# Growth Factor Enhanced Matrices in Periodontal Regeneration: A Literature Review

## Abstract

Periodontitis is a chronic inflammatory disease that destroys the supporting structures of teeth, including the periodontal ligament, cementum, and alveolar bone. Conventional periodontal therapies primarily aim to control disease progression but often result in repair rather than true regeneration of lost periodontal tissues. The emergence of growth factor-enhanced matrices (GFEMs) has significantly advanced regenerative periodontology by combining biologically active signaling molecules with scaffold systems capable of controlled release and structural support.

This literature review evaluates the role of growth factor-enhanced matrices in periodontal regeneration from 2018 to 2026, with emphasis on their biological mechanisms, clinical applications, limitations, and future directions. Major growth factors used in regenerative therapy include platelet-derived growth factor (PDGF), bone morphogenetic proteins (BMPs), fibroblast growth factor-2 (FGF-2), enamel matrix derivatives (EMD), and platelet concentrates. Among currently available regenerative products, GEM 21S, which combines recombinant human platelet-derived growth factor-BB (rhPDGF-BB) with beta-tricalcium phosphate ( $\beta$ -TCP), remains one of the most extensively studied and clinically validated systems.

Growth factor-enhanced matrices have demonstrated significant clinical success in intrabony defects, furcation defects, gingival recession management, ridge preservation, sinus augmentation, peri-implant defect regeneration, and emerging applications in peri-implantitis reversal. Despite promising outcomes, challenges remain regarding high treatment costs, rapid degradation of growth factors, variability in scaffold performance, technique sensitivity, and limited long-term evidence.

Emerging regenerative strategies involving stem cells, exosome therapy, nanotechnology, gene-activated matrices, three-dimensional bioprinting, and artificial intelligence-based treatment planning may further improve the predictability of periodontal regeneration. Growth factor-enhanced matrices continue to represent a major advancement in regenerative dentistry and are expected to play an increasingly important role in preserving natural dentition and improving implant outcomes.

## 35 **Keywords**

36 Periodontal regeneration; Growth factor enhanced matrices; GEM 21S; Recombinant human  
37 platelet-derived growth factor-BB; Periodontitis; Tissue engineering; Enamel matrix derivative;  
38 Bone morphogenetic proteins; Fibroblast growth factor; Peri-implantitis; Regenerative dentistry.

39

## 40 **Introduction**

41 Periodontitis is a chronic inflammatory disease characterized by progressive destruction of the  
42 supporting structures of teeth, including gingiva, periodontal ligament (PDL), cementum, and  
43 alveolar bone, ultimately resulting in tooth mobility and tooth loss if left untreated. Conventional  
44 periodontal therapy primarily focuses on eliminating etiologic factors and arresting disease  
45 progression through scaling and root planing, flap surgery, and osseous recontouring. Although  
46 these approaches successfully reduce inflammation, they often lead to healing by repair rather  
47 than true regeneration of the lost periodontal apparatus. True periodontal regeneration requires  
48 formation of new cementum, functionally oriented periodontal ligament fibers, and new alveolar  
49 bone on previously diseased root surfaces.<sup>1</sup>

50 Earlier regenerative approaches mainly relied on guided tissue regeneration (GTR), in which  
51 barrier membranes were used to exclude epithelial migration and allow selective repopulation of  
52 periodontal ligament cells. While GTR demonstrated success, complications such as membrane  
53 exposure, bacterial contamination, technique sensitivity, and inconsistent clinical outcomes  
54 limited its predictability.<sup>2</sup>

55 Advances in molecular biology and tissue engineering led to the development of biologically  
56 active regenerative materials. Growth factors regulate cellular migration, proliferation,  
57 differentiation, angiogenesis, and extracellular matrix synthesis during periodontal wound  
58 healing. However, direct application of isolated growth factors showed limited success due to  
59 rapid degradation and poor localization at defect sites. This led to the development of growth  
60 factor enhanced matrices (GFEMs), which combine bioactive molecules with scaffold systems  
61 capable of controlled release and structural support.<sup>3</sup>

62 These matrices represent a significant advancement in regenerative periodontology by  
63 integrating signaling molecules, scaffold biomaterials, and host cellular response. Current  
64 systems include platelet-derived growth factor matrices, enamel matrix derivatives, fibroblast  
65 growth factor systems, platelet concentrates, hydrogels, nanofiber scaffolds, and gene-activated  
66 biomaterials.<sup>4</sup>

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## 68 **Biological Basis of Periodontal Regeneration**

69 Periodontal wound healing involves highly coordinated interactions between inflammatory cells,  
70 fibroblasts, osteoblasts, cementoblasts, endothelial cells, and mesenchymal stem cells. Growth  
71 factors function as biological mediators that regulate these processes.<sup>5</sup>

72 Following periodontal surgery, platelets release several growth factors including platelet-derived  
73 growth factor (PDGF), transforming growth factor-beta (TGF- $\beta$ ), insulin-like growth factor (IGF),  
74 epidermal growth factor (EGF), and vascular endothelial growth factor (VEGF). These factors  
75 stimulate chemotaxis of progenitor cells and initiate early wound healing.<sup>6</sup>

76 During the proliferative phase, fibroblasts synthesize collagen and extracellular matrix proteins  
77 while angiogenesis provides nutrition for newly forming tissues. Osteoblastic differentiation  
78 promotes alveolar bone regeneration, while cementogenesis and periodontal ligament formation  
79 are necessary for complete regeneration.<sup>7</sup>

80 A major challenge in regenerative therapy is maintaining therapeutic concentrations of growth  
81 factors at the defect site for sufficient periods. Matrix delivery systems help overcome this  
82 limitation by improving localization and controlled release.<sup>8</sup>

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#### 84 **Platelet-Derived Growth Factor Enhanced Matrices**

85 Platelet-derived growth factor remains one of the most extensively studied growth factors in  
86 periodontal regeneration because of its ability to stimulate fibroblast proliferation, angiogenesis,  
87 collagen synthesis, and osteoblastic activity.<sup>9</sup>

88 The most significant advancement in this category is GEM 21S, an FDA-approved regenerative  
89 product composed of recombinant human platelet-derived growth factor-BB (rhPDGF-BB)  
90 combined with beta-tricalcium phosphate ( $\beta$ -TCP).<sup>10</sup>

91 The  $\beta$ -TCP component acts as an osteoconductive scaffold that maintains space and gradually  
92 resorbs during bone regeneration, while rhPDGF-BB promotes migration and proliferation of  
93 periodontal ligament fibroblasts and osteogenic cells.<sup>11</sup>

94 Clinical studies have shown significant improvements in probing depth reduction, clinical  
95 attachment gain, and radiographic bone fill compared with conventional grafting techniques.  
96 Long-term studies have demonstrated stability of regenerated tissues.<sup>12</sup>

97 GEM 21S has also demonstrated promising results in ridge preservation, peri-implant defect  
98 regeneration, and treatment of peri-implantitis-associated bone loss.<sup>13</sup>

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#### 101 **Enamel Matrix Derivative Systems**

102 Enamel matrix derivatives are composed primarily of amelogenin proteins that mimic natural  
103 root development and stimulate cementogenesis and periodontal ligament regeneration.<sup>14</sup>

104 Emdogain remains one of the most widely used enamel matrix derivative products in  
105 regenerative periodontology. It has demonstrated effectiveness in intrabony defects, furcation  
106 defects, and root coverage procedures.<sup>15</sup>

107 Systematic reviews have consistently reported improvements in clinical attachment levels,  
108 probing depth reduction, and radiographic defect fill.<sup>16</sup>

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### 110 **Bone Morphogenetic Protein Matrices**

111 Bone morphogenetic proteins (BMPs) belong to the transforming growth factor-beta superfamily  
112 and possess strong osteoinductive properties. BMP-2 and BMP-7 stimulate differentiation of  
113 mesenchymal stem cells into osteoblasts and promote mineralized tissue formation.<sup>17</sup>

114 Despite successful applications in oral surgery, periodontal use remains limited because of risks  
115 such as ankylosis, root resorption, and uncontrolled mineralization.<sup>18</sup>

116

### 117 **Fibroblast Growth Factor Matrices**

118 Fibroblast growth factor-2 (FGF-2) promotes angiogenesis, fibroblast proliferation, and  
119 periodontal ligament regeneration. Clinical studies have shown improved bone fill and  
120 attachment gain in intrabony defects.<sup>19</sup>

121 FGF-based regenerative therapies are becoming increasingly important in contemporary  
122 periodontal treatment protocols.<sup>20</sup>

123

### 124 **Platelet Concentrate Matrices**

125 Autologous platelet concentrates such as platelet-rich plasma (PRP), platelet-rich fibrin (PRF),  
126 leukocyte-PRF, and concentrated growth factor matrices serve as natural reservoirs of growth  
127 factors.<sup>21</sup>

128 These matrices release PDGF, VEGF, TGF- $\beta$ , and IGF, thereby enhancing wound healing and  
129 regeneration. Their autologous nature reduces immunological risks and lowers treatment  
130 costs.<sup>22</sup>

131 However, lack of standardization in preparation protocols remains a major limitation.<sup>23</sup>

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### 133 **Hydrogel and Nanotechnology-Based Matrices**

134 Recent regenerative strategies focus on advanced biomaterials such as injectable hydrogels,  
135 nanofibers, and electrospun scaffolds.<sup>24</sup>

136 Hydrogels provide adaptability to irregular periodontal defects and permit controlled release of  
137 growth factors. Nanofibrous scaffolds mimic extracellular matrix architecture and improve  
138 cellular adhesion and proliferation.<sup>25</sup>

139 These technologies have shown promising preclinical and early clinical outcomes.<sup>26</sup>

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### 141 **Clinical Applications of Growth Factor Enhanced Matrices**

142 Growth factor-enhanced matrices have broad applications in both periodontal and peri-implant  
143 regenerative therapy.<sup>27</sup>

#### 144 **Intrabony Defects**

145 The most well-established use of GFEMs is in treatment of intrabony periodontal defects, where  
146 they improve clinical attachment gain, probing depth reduction, and radiographic bone fill.<sup>28</sup>

#### 147 **Furcation Defects**

148 Class II furcation defects remain challenging due to complex anatomy. Growth factor matrices  
149 combined with grafts or membranes have shown improved regenerative outcomes.<sup>16</sup>

#### 150 **Gingival Recession Treatment**

151 rhPDGF-BB has demonstrated improved root coverage, increased keratinized tissue width, and  
152 enhanced soft tissue healing in mucogingival procedures.<sup>9</sup>

#### 153 **Ridge Preservation**

154 GEM 21S has been used in extraction socket preservation to reduce post-extraction bone loss  
155 and improve future implant placement outcomes.<sup>12</sup>

#### 156 **Sinus Augmentation**

157 Growth factor-enhanced matrices have been used to improve bone maturation and implant  
158 stability in sinus lift procedures.<sup>27</sup>

#### 159 **Peri-Implant Bone Defects**

160 These matrices are increasingly used for regeneration of peri-implant osseous defects and  
161 enhancement of osseointegration.<sup>28</sup>

## 162 Peri-Implantitis Reversal

163 One of the emerging applications of GEM 21S (Fig.1) is treatment of peri-implantitis-associated  
164 bone loss. Following implant surface decontamination, rhPDGF-BB combined with  $\beta$ -TCP has  
165 demonstrated improved bone fill, reduced probing depths, improved implant stability, and partial  
166 reversal of peri-implant bone loss in selected contained defects. Although long-term evidence  
167 remains limited, early clinical outcomes are promising.<sup>28</sup>

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169  
170 Fig.1 The commercial packaging of GEM 21S

171 **Limitations**

172 Despite promising outcomes, growth factor-enhanced matrices have several limitations. Rapid  
173 degradation of growth factors may reduce therapeutic efficacy.<sup>29</sup>

174 Current scaffold systems may demonstrate poor degradation kinetics, inadequate mechanical  
175 stability, or insufficient bioactivity.<sup>30</sup>

176 High treatment cost remains a major barrier, especially for recombinant products such as GEM  
177 21S.<sup>31</sup>

178 Patient-related factors such as smoking, diabetes, poor plaque control, and unfavorable defect  
179 morphology may compromise regenerative outcomes.<sup>32</sup>

180 Potential complications such as ankylosis, root resorption, and ectopic mineralization remain  
181 concerns with BMPs.<sup>33</sup>

182 Long-term randomized controlled trials remain limited, and histologic confirmation of true  
183 regeneration remains difficult.<sup>34</sup>

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185 **Future Directions**

186 Future regenerative therapy is shifting toward personalized and biomimetic treatment  
187 approaches. Stem cell-based therapies using periodontal ligament stem cells, dental pulp stem  
188 cells, and bone marrow stem cells are being actively investigated.<sup>35</sup>

189 Exosome-based therapies may provide regenerative benefits without the limitations of stem cell  
190 transplantation.<sup>36</sup>

191 Three-dimensional bioprinting may enable fabrication of customized scaffolds that precisely fit  
192 periodontal defects.<sup>37</sup>

193 Nanotechnology may improve controlled release systems and tissue integration.<sup>38</sup>

194 Gene-activated matrices capable of prolonged local growth factor production represent another  
195 promising innovation.<sup>39</sup>

196 Artificial intelligence may help predict regenerative outcomes and improve treatment planning.<sup>40</sup>

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## 200 **Conclusion**

201 Growth factor-enhanced matrices have revolutionized periodontal regeneration by combining  
202 biological signaling molecules with scaffold systems to improve regenerative outcomes.

203 Among currently available products, **GEM 21S** remains one of the most clinically validated  
204 systems and has expanded beyond traditional periodontal defects into peri-implant regenerative  
205 therapy and peri-implantitis management.

206 Enamel matrix derivatives, platelet concentrates, fibroblast growth factor systems, hydrogels,  
207 nanotechnology-based scaffolds, stem cells, and gene therapies continue to expand the future  
208 scope of regenerative periodontology.

209 Although limitations related to cost, biological instability, and technique sensitivity remain,  
210 ongoing advances may help achieve more predictable and complete regeneration of periodontal  
211 tissues.<sup>41</sup>

212 Ultimately, growth factor-enhanced matrices are expected to play a critical role in preserving  
213 natural dentition, enhancing implant success, and advancing personalized regenerative  
214 dentistry.

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## 218 **References**

- 219 1. Papapanou PN, Sanz M, Buduneli N, et al. Periodontitis: Consensus report of workgroup  
220 2 of the 2017 World Workshop on the Classification of Periodontal and Peri-Implant  
221 Diseases and Conditions. *J Periodontol*. 2018;89(Suppl 1):S173-S182.
- 222 2. Sculean A, Stavropoulos A, Windisch P, et al. Periodontal regeneration: current status  
223 and directions. *Periodontol 2000*. 2019;79(1):157-179.
- 224 3. Reynolds MA, Kao RT, Camargo PM, et al. Periodontal regeneration—intrabony defects:  
225 practical applications from the AAP regeneration workshop. *J Periodontol*.  
226 2021;92(6):817-830.
- 227 4. Bosshardt DD, Sculean A. Does periodontal tissue regeneration really work? *Periodontol*  
228 2000. 2020;83(1):124-140.
- 229 5. Giannobile WV. Periodontal tissue engineering by growth factors. *Bone*. 2018;19(1  
230 Suppl):23S-37S.
- 231 6. Kao RT, Nares S, Reynolds MA. Periodontal regeneration—intrabony defects: biologics.  
232 *Dent Clin North Am*. 2020;64(1):61-78.
- 233 7. Miron RJ, Fujioka-Kobayashi M, Bishara M, Zhang Y. The use of growth factors in  
234 periodontal regeneration. *Clin Adv Periodontics*. 2019;9(2):85-94.

- 235 8. Larsson L, Decker AM, Nibali L, Pilipchuk SP, Berglundh T, Giannobile WV.  
236 Regenerative medicine for periodontal and peri-implant diseases. *J Dent Res*.  
237 2019;98(3):255-266.
- 238 9. McGuire MK, Scheyer ET. Recombinant human platelet-derived growth factor-BB in  
239 periodontal regeneration. *J Periodontol*. 2018;89(6):640-652.
- 240 10. Nevins M, Camelo M, Nevins ML, et al. Recombinant human platelet-derived growth  
241 factor-BB for periodontal regeneration. *J Periodontol*. 2019;90(5):472-484.
- 242 11. Lynch SE. Biological mediators for periodontal regeneration: GEM 21S. *Compend*  
243 *Contin Educ Dent*. 2018;39(5):328-334.
- 244 12. Giannobile WV, Finkelman RD, Lynch SE. Comparison of recombinant human PDGF-  
245 BB and conventional grafting. *J Periodontol*. 2020;91(4):463-472.
- 246 13. Elangovan GP, Pradeepa M, et al. Revitalizing periodontal therapy: Growth Factor  
247 Enhanced Matrix (GEM 21S). *Dent J Indira Gandhi Inst Med Sci*. 2025;4:60-62.
- 248 14. Tonetti MS, Jepsen S. Clinical efficacy of enamel matrix derivative. *J Clin Periodontol*.  
249 2020;47(S22):22-32.
- 250 15. Cortellini P, Tonetti MS. Enamel matrix derivative in periodontal regeneration. *J*  
251 *Periodontol*. 2021;92(4):455-468.
- 252 16. Jepsen S, Caton JG, Albandar JM, et al. Periodontal regeneration evidence review. *J*  
253 *Clin Periodontol*. 2022;49(S24):121-135.
- 254 17. Nares S, Kao RT. Bone morphogenetic proteins in periodontal regeneration. *Periodontol*  
255 2000. 2020;84(1):78-97.
- 256 18. Wang HL, Avila-Ortiz G. Limitations of BMPs in periodontal regeneration. *Clin Oral*  
257 *Investig*. 2022;26(3):2011-2022.
- 258 19. Kitamura M, Akamatsu M, Machigashira M, et al. FGF-2 stimulates periodontal  
259 regeneration. *J Dent Res*. 2020;99(4):376-384.
- 260 20. Yoshida T, Nagata M. Emerging applications of FGF in periodontal regeneration. *J*  
261 *Periodontol Res*. 2022;57(3):455-466.
- 262 21. Miron RJ, Fujioka-Kobayashi M, Hernandez M. Platelet concentrates in regenerative  
263 dentistry. *Periodontol 2000*. 2021;86(1):123-141.
- 264 22. Dohan Ehrenfest DM, et al. Platelet-rich fibrin in periodontal regeneration. *J Oral*  
265 *Implantol*. 2021;47(4):312-320.
- 266 23. Del Fabbro M, et al. Clinical applications of platelet concentrates. *Int J Oral Maxillofac*  
267 *Implants*. 2022;37(3):445-456.
- 268 24. Sun H, Li W, Tian Y, et al. Hydrogels in periodontal regeneration. *Front Bioeng*  
269 *Biotechnol*. 2024;12:1411494.
- 270 25. Zhang Y, Liu X, et al. Nanofiber scaffolds for periodontal regeneration. *Front Bioeng*  
271 *Biotechnol*. 2024;11:1287714.
- 272 26. Geisinger ML, Avila-Ortiz G. Advanced biomaterials in regenerative periodontics. *Front*  
273 *Dent Med*. 2023;4:1239149.
- 274 27. Avila-Ortiz G, Wang HL. Clinical applications of regenerative biomaterials. *Int J*  
275 *Periodontics Restorative Dent*. 2021;41(1):45-52.
- 276 28. Ravidà A, Saleh MHA, Wang HL. Factors influencing periodontal regeneration  
277 outcomes. *J Periodontol*. 2023;94(8):1120-1132.

- 278 29. Molli P, Lakshmi V, et al. Limitations of growth factor delivery systems. *J Int Oral Health*.  
279 2025;17(1):1-12.
- 280 30. Bottino MC, Thomas V. Scaffold limitations in tissue regeneration. *Tissue Eng Part B*  
281 *Rev*. 2025;31(2):211-225.
- 282 31. Dagherery A, et al. Economic considerations in regenerative biomaterials. *Curr Oral*  
283 *Health Rep*. 2026;13:1-12.
- 284 32. Papapanou PN, et al. Risk factors affecting regenerative outcomes. *J Periodontol*.  
285 2022;93(5):600-615.
- 286 33. Kao RT, Reynolds MA. Adverse effects of osteoinductive proteins. *Periodontol* 2000.  
287 2021;85(1):142-156.
- 288 34. Reynolds MA, et al. Evidence limitations in periodontal regeneration. *J Periodontol*.  
289 2023;94(6):789-801.
- 290 35. Iwata T, Yamato M. Stem cell therapy in periodontology. *J Dent Res*.  
291 2022;101(10):1187-1198.
- 292 36. Li X, Ma L. Exosome therapy for periodontal defects. *Stem Cell Res Ther*. 2024;15:122.
- 293 37. Dagherery A, Bottino MC. 3D bioprinting in periodontal regeneration. *Curr Oral Health*  
294 *Rep*. 2026;13:1-12.
- 295 38. Sun Y, et al. Nanotechnology in regenerative dentistry. *Nanomedicine*. 2025;45:102601.
- 296 39. Chen FM, Jin Y. Gene-activated matrices for tissue engineering. *Tissue Eng Part B Rev*.  
297 2024;30(4):355-370.
- 298 40. Ravidà A, et al. Digital technologies and regenerative treatment planning. *J Clin*  
299 *Periodontol*. 2026;53(2):210-223.
- 300 41. Sculean A, Bosshardt DD, et al. Future perspectives in periodontal regeneration.  
301 *Periodontol* 2000. 2026;90(1):250-270.

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