

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30

Association between Paraoxonase 1 Allelic Frequency Distribution, Adjusted Odds Ratios, and the Risk of Cardiovascular Diseases in a Moroccan population.

ABSTRACT:

Aim : Paraoxonase 1 (PON1) an antioxidant enzyme associated with high density lipoprotein (HDL) is expressed in the Islet of Langerhans and may play a protective role against oxidative stress. This study investigated the relationship between the PON1 phenotype, allelic frequency distribution and the risk of cardiovascular disease complications.

Methods : Three hundred subjects aged between 40-80 years including healthy controls, diabetic patients, and chronic renal failure/hemodialysis patients, were enrolled and divided into three groups of 100 subjects each. Participants were further stratified by age into middle-aged 40-59 years, (n=147), and elderly subjects 60-79 years (n = 153).

Plasma malondialdehyde levels were measured by HPLC using thiobarbituric acid (TBA) derivatization with fluorescence detection. Vitamin E (α -tocopherol) concentration was determined by HPLC with electrochemical detection.

Results : PON 1 allelic frequencies in healthy, diabetic and hemodialysis subjects were as respectively : Q alleles 86.62%, 85.46% and 79.78%; R alleles 13.37%, 14.53% and 20.22%. In hemodialysis patients, the adjusted odds ratio (OR) comparing the QQ with the RR phenotypes was 1.97 [95 % (CI): 0.63-6.21]. In diabetic patients, the OR comparing the QQ with the QR phenotypes was 1.37 [95% CI: 0.62-3.04]. PON1 activity was significantly higher in QR+RR carriers than in QQ carriers among diabetic and hemodialysis patients ($p<0.001$ for both groups). The vitamin E/Total Cholesterol ratio was significantly reduced in diabetic QQ carriers ($p<0.05$) and hemodialysis QQ carriers ($p<0.05$) compared with healthy subjects.

Conclusion : These finding suggest an association between changes in PON1 phenotype distribution, allelic frequencies, and the development of cardiovascular complication in diabetic and hemodialysis patients.

31

32

33 **Keywords** : Adjusted odd ratio, Paraoxonase 1, HPLC, Hemodialysis, Diabetes, MDA,
34 Vitamin E

35

36 **Introduction**

37

38 Diabetes and renal failure are increasingly prevalent health problems in developing countries,
39 including Morocco and represent major risk factors for cardiovascular disease and myocardial
40 infarction [1-5]. Paraoxonase 1 (PON1) is an antioxidant enzyme associated with HDL that
41 hydrolyzes lipid peroxides in LDL and atherosclerotic lesions [6-7]. Three paraoxonase
42 isoforms have been identified (PON1, PON2 and PON3), although only PON1 exhibits true
43 paraoxonase activity. Oxidized LDL plays a central role in the cardiovascular complications
44 associated with diabetes and hemodialysis [7]. Reduced PON1 activity in diabetic and
45 hemodialysis patients may contribute to increased susceptibility to lipid peroxidation, partly due
46 to hyperglycaemia and the dialysis process [5-12]. Therefore, elevated paraoxonase activity
47 may provide protective cardiovascular effects in these populations [2,13]. One of the factor
48 determinants of PON1 activity is the Q192R genetic polymorphism, which give rises to two
49 isoforms: Q (low Paraoxonase activity) and R (high Paraoxonase activity) [8, 9]. The R allele
50 has been associated with increased risk factor for cardiovascular disease and diabetic or
51 hemodialysis patients carrying this allele may be at greater risk of cardiovascular
52 complications [2, 7-9].

53 Previously, we determined the distribution of PON1 phenotypes in a Moroccan population
54 [2]. The present study aimed to investigate the association between alterations in PON1
55 phenotype distribution and the risk of cardiovascular disease in healthy subjects, diabetic
56 patients, and hemodialysis patients.

57

58

59

60

61

62

63

64

65 **1-Materials and methods**

66 **1-1 Subjects**

67 Three hundred subjects aged 40-80 years were enrolled in this study. Participants were
68 recruited from the Biomedical center of the Pasteur Institute of Casablanca for routine
69 medical evaluation. Diabetic patients (n = 100) were recruited from local clinical centers,
70 whereas hemodialysis patients (n = 100) were recruited from hemodialysis centers in
71 Casablanca. Participants were divided into three group according to health status: healthy
72 control (n=100), diabetic patients (n = 100), hemodialysis patients (n = 100). Pregnant and
73 breastfeeding women were excluded, and only one individual per family was included among
74 diabetic and hemodialysis patients.

75 Healthy subjects were nonsmokers and nondrinkers with no clinically apparent
76 cardiovascular disease, normal renal and liver function and no clinical or laboratory evidence
77 of diabetes or inflammation (hs-CRP < 3; fasting blood glucose < 6.1 mmol/l). None were
78 taking medication or oral antioxidant supplements.

79 Among diabetic patients, 75 % were hypertensive, 23 % had a family history of
80 diabetes, and most were receiving statin therapy. The mean duration of was 4.33 ± 2.42 years,
81 and diabetic nephropathy was present in 22% of patients. One subject had previously
82 undergone surgery for disc herniation, and another had a solitary kidney. Cardiovascular
83 disease was present in 40 % of diabetic patients, and three patients were treated with insulin.

84 Hemodialysis subjects received standard unfractionated heparin administered in
85 fractional doses every hour during dialysis sessions. The delivered dialysis dose corresponded
86 to a Kt/V is 1.2. Patients underwent hemodialysis for 4 hours, three times per week, using
87 bicarbonate-based dialysate. Hemophane membranes were the most used membranes. Among
88 the hemodialysis subjects, 55 % were hypertensive and 15 % were receiving statin therapy.

89 The mean duration of hemodialysis treatment was 53 months. One subject had previously
90 undergone kidney transplantation, another had valvular heart disease, and one subject had a
91 solitary kidney. The study protocol was accepted by the ethics committee of the Faculty of
92 Medicine and Pharmacy of Casablanca, and all participants provided written informed
93 consent prior to enrollment.

94 All subjects underwent a physical examination and completed a self-administered
95 questionnaire. This questionnaire collected information on demographic characteristics,
96 participants' health status, family history of diabetes, hypertension and cardiovascular
97 diseases. It also included information regarding the duration of diabetes and hemodialysis
98 treatment.

99 Body mass index (BMI) was calculated from weight and Height measurements. Blood pressure
100 was measured twice on the right arm in supine position after 5-minutes rest period to
101 determine hypertension according to the 1999 World Health Organisation criteria.
102 Measurement were performed under quiet conditions and in the absence of emotional stress.

103

104 **1-2 Blood and urine collection and lipid profile measurements**

105

106 After an overnight fast, 80 mL of blood were collected into sodium heparin- or EDTA-
107 containing tubes. Plasma was separated by centrifugation (3000 x g, for 10 min) and serum
108 aliquots were stored at -80 °C until analysis. Microalbuminuria was in two consecutive 24-h
109 urine samples using a BN 100 nephelometer system (Dade Behring, Germany). Serum urea
110 and creatinine levels were assessed before each dialysis session in hemodialysis patients.
111 Serum total cholesterol, HDL cholesterol, LDL cholesterol, triglycerides, and HbA1c were
112 measured using automated enzymatic assays (Kodak, Ektachem USA System). Serum total
113 cholesterol levels were determined by measuring absorbance at 550 nm.

114

115

116 **1.3. Paraoxonase activity**

117

118 PON1 activity was determined using paraoxon (O,O-diethyl-O-p-nitrophenylphosphate;
119 Sigma) as the substrate by measuring the increase in absorbance at 412 nm resulting from the
120 formation of 4-nitrophenol, as previously described [2]. Briefly, 50 µl of serum were added
121 to 1 ml of Tris-HCl buffer (100 mmol/l, pH 8.0) containing 2 mmol/l CaCl₂ and 5.5 mmol/l
122 paraoxon [9, 11, 14, 15], and the reaction was monitored at 25 °C. Enzymatic activity was

123 calculated using a molar extinction coefficient $17,100 \text{ M}^{-1} \text{ cm}^{-1}$. One unit of paraoxonase
124 activity was defined as 1 nmol of 4-nitrophenol generated per minute under the assay
125 conditions.

126

127

128

129

130

131 **1.4. Arylesterase assay**

132 Arylesterase activity was measured spectrophotometrically using phenylacetate as
133 substrate in 20 mM Tris/HCl pH 8.0 [16]. The increase in absorbance was monitored at 270
134 nm, and enzyme activity was calculated using a molar extinction coefficient of $1310 \text{ M}^{-1} \text{ cm}^{-1}$.
135 One unit (U) corresponded to 1 μmol phenylacetate hydrolyzed per minute [6, 16].

136

137

138 **1.5. Paraoxonase phenotype distribution**

139 The PON1 Q192R polymorphism determines two isoforms: Q (low activity) and R (high
140 activity) [8]. PON1 phenotype was determined using the dual substrate method based on the
141 ratio of salt-stimulated paraoxonase activity (1 mol/l NaCl) to arylesterase activity. Subjects
142 were classified as QQ (ratio < 3.0), QR (ratio 3.0-7) and ratio > 7.0 , or RR phenotype (ratio $>$
143 7.0) [10, 11].

144

145 **1.6. Statistical analysis**

146 Statistical analysis were performed using SAS for windows version 6.11. Data are presented
147 as mean \pm standard deviation of number cases. Comparisons between groups were
148 performed using unpaired t-tests or ANOVA. Associations between variables were assessed
149 using regression and Spearman correlation analyses. A two sided p value < 0.05 was
150 considered statistically significant.

151

152 **2. Results**

153 **2.1. Baseline data**

154 In the overall study population, MDA concentrations tended to be higher in carriers of the
155 PON1 QR + RR phenotypes compared with carriers of the PON1 QQ phenotype ($p = 0.08$).

156 MDA concentrations were also higher in PON1 QR carriers than in PON1 RR carriers ($p =$
157 0.08). In contrast, subjects with the PON1 QR phenotype had significantly higher MDA
158 concentrations than PON1 QQ carriers ($p < 0.05$) (Table 1).

159 Among PON1 QQ carriers, MDA levels were significantly higher in diabetic subjects
160 ($p < 0.05$) and hemodialysis subjects ($p < 0.01$) compared with healthy subjects with the same
161 phenotype. Similarly, diabetic and hemodialysis subjects carrying the PON1 QR phenotype
162 had higher MDA levels than healthy PON1 QR carriers ($p < 0.05$). Among PON1 RR carriers,
163 MDA levels were increased in diabetic ($p = 0.32$) and hemodialysis subjects ($p = 0.33$)
164 compared with healthy RR carriers, but these differences were not statistically significant.

165 Paraoxonase activity was significantly higher in subjects carrying the PON1 QR or RR
166 phenotypes compared with PON1 QQ carriers ($p < 0.001$). Indeed, arylesterase activity was
167 significantly lower in carriers of the PON1 R allele (RR and QR) compared with carriers of
168 the PON1 Q allele ($p < 0.01$). Furthermore, arylesterase activity was significantly lower in
169 PON1 RR carriers than in PON1 QR carriers ($p < 0.01$) and in combined PON1 QR+RR
170 carriers ($p < 0.05$). Consistently, arylesterase activity was significantly higher in PON1 QQ
171 carriers compared with combined PON1 QR + RR carriers ($p < 0.001$) (Table 1).

172 Paraoxonase ($p < 0.05$) and arylesterase ($p < 0.05$) activities were significantly lower in the
173 hemodialysis subjects across all three PON1 Q192R phenotypes compared with healthy
174 subjects. In contrast, non-significant differences in paraoxonase activity were observed
175 between healthy subjects and the diabetic patients for any of the three PON1 Q192R
176 phenotypes, although a slight increase in serum PON1 activity was noted in diabetic patients.
177 Paraoxonase ($p < 0.05$) and Arylesterase ($p < 0.01$) activities also differed significantly among
178 the three PON1 Q192R phenotypes in both the diabetic and hemodialysis subjects. In healthy
179 subjects, carriers of the PON1 QQ phenotype had significantly lower paraoxonase activity
180 compared with QR + RR carriers ($p < 0.001$). Similarly, paraoxonase activity was significantly
181 higher in QR+RR carriers than in QQ carriers among diabetic ($p < 0.001$) and hemodialysis
182 patients ($p < 0.001$).

183 Comparison of Vitamin E/Total cholesterol ratio in the overall study population according
184 to the PON1 Q192R polymorphism showed a significant decrease in PON1 QR carriers when
185 compared with PON1 QQ carriers ($p < 0.05$). In contrast, the Vitamin E/total cholesterol ration
186 was significantly lower in PON1 QQ carriers compared with combined PON1 QR + RR
187 carriers ($p < 0.05$). However, no significant difference in the vitamin E/total cholesterol ratio
188 was observed between PON1 QQ and PON1 RR carriers ($p = 0.10$) (Table 1).

189 Vitamin E levels did not differ significantly among subjects according to the PON1 Q192R
 190 polymorphism; however, the Vitamin E/total cholesterol was significantly influenced by the
 191 PON1 phenotype (Table 1). Compared with healthy subjects carrying the PON1 QQ
 192 phenotype, the Vitamin E/total cholesterol ratio was significantly lower in diabetic ($p<0.05$)
 193 and hemodialysis subjects ($p<0.05$) carrying the same phenotype.

194 A significant decrease in the Vitamin E/total cholesterol ratio was observed in hemodialysis
 195 subjects carrying the PON1 QR phenotype compared with healthy QR carriers ($p<0.05$).
 196 Similarly, hemodialysis subjects carrying the PON1 RR phenotype had a significantly lower
 197 ratio Vitamin E/total cholesterol ratio than the healthy RR carriers ($p < 0.05$). Interestingly,
 198 the vitamin E/total cholesterol ratio was significantly decreased in hemodialysis subjects
 199 carrying the combined PON1 QR + RR phenotypes compared with healthy QR+RR carriers
 200 ($p<0.01$).

201 However, comparisons between PON1 QQ carriers and combined PON1 QR + RR carriers
 202 within healthy ($p = 0.14$), diabetic ($p = 0.32$), and hemodialysis subjects ($p = 0.10$) showed no
 203 significant differences in the Vitamin E/total cholesterol ratio.

204

205 **Table 1 : Baseline characteristics (mean \pm SD) according to PON1 Q/R in**
 206 **whole subjects (n=300)**

Parameters	QQ (n=195)	QR (n=48)	QR+ RR (n=66)	RR (n=18)
Men (%)	58.46	46.93	46.97	47.05
Mean age, years	57.82 \pm 11.59	57.66 \pm 11.72	57.30 \pm 11.82	56.33 \pm 12.38
GJ (g/L)	1.50 \pm 1.00	1.52 \pm 0.60	1.53 \pm 0.64	1.57 \pm 0.83
TG, (mmol/L)	1.50 \pm 1.04	2.34 \pm 1.75 ^{***}	2.33 \pm 1.74 ^{***}	1.37 \pm 0.55
Serum MDA (μ M)	7.43 \pm 3.83	8.76 \pm 4.61 [*]	8.26 \pm 4.36	7.04 \pm 3.48
PON1 activity (U/mL)	38.68 \pm 47.95	110.45 \pm 119.40 ^{***}	113.67 \pm 131.35 ^{***}	118.09 \pm 147.87 ^{***}
ARE activity (U/mL)	38.28 \pm 34.24	23.72 \pm 25.21 ^{**}	19.47 \pm 23.02 ^{***¶}	8.14 \pm 9.14 ^{**++}
Vit. E (μ M)	18.67 \pm 8.52	18.66 \pm 7.13	18.57 \pm 7.63	18.40 \pm 8.93
Vit.E/TC ratio (μ mol/mmol)	4.66 \pm 2.42	3.78 \pm 1.83 [*]	3.79 \pm 1.95 [*]	3.80 \pm 2.21

207

208 *Values are mean ± SD., unless indicated otherwise. The unpaired student ttest was applied.*
209 *Significance was calculated in comparison to QQ, QR, QR + RR, RR carriers : *p<0.05, **p<0.01,*
210 ****p<0.001 and in comparisons between QQ and RR carriers ++p<0.01, and in comparison*
211 *between QQ and QR+ RR carriers †p<0.05*

212 In diabetic patients, the adjusted OR comparing the PON1 QQ variant with the PON1 QR
213 variant was 1.37 (95% CI: 0.62-3.04). In contrast, the adjusted OR comparing the PON1 QQ
214 and PON1 RR variants was 0.84 (95% CI: 0.22-3.25). Among hemodialysis patients, the
215 adjusted ORs comparing the PON1 QQ variant with the PON1 QR and PON1 RR variants
216 were 1.52 (95% CI: 0.69-3.35) and 1.97 (95 % CI: 0.63-6.21).

217 The observed allelic frequency distribution of PON1 showed Q allele frequencies of 86.62%,
218 85.46%, and 79.78%, for R allele frequencies of 13.37%, 14.53%, and 20.22% in healthy,
219 diabetic and hemodialysis patients, respectively.

220 The adjusted ratio OR1 comparing the frequency of the PON1 risk allele R between healthy
221 and diabetic subjects was 1.10 (95% CI: 0.492.45). Similarly, the OR2 comparing healthy and
222 hemodialysis subjects was 1.64 (95% CI: 0.773.49) (Table 2).

223 Our result showed that, triglycerides levels were significantly higher in hemodialysis patients
224 carrying the PON1 QQ phenotype compared with healthy QQ carriers (p<0.001). Similarly,
225 triglyceride concentrations were significantly elevated in hemodialysis patients carrying the
226 PON1 QR (p < 0.001) and combined PON1 QR + RR phenotypes (p<0.001) compared with
227 healthy subjects with the corresponding phenotypes.

228 Higher triglyceride concentrations were also observed in diabetic subjects carrying the PON1
229 QQ phenotype compared with healthy QQ carriers, although the difference was not
230 statistically significant (p = 0.12). A similar trend was observed in diabetic subjects carrying
231 the combined PON1 QR + RR phenotypes compared with healthy QR+RR carriers (p = 0.13).
232 In contrast, diabetic patients carrying the PON1 QR phenotype had significantly higher
233 triglyceride levels than healthy QR carriers (p< 0.05).

234 Interestingly, hemodialysis exhibited significantly higher triglyceride levels than diabetic
235 subjects across the PON1 QQ (p < 0.001), PON1 QR (p < 0.001), combined PON1 QR + RR
236 phenotype (p<0.001).

237

238

239

240

241

242

243

244

245

Allelic frequency	Healthy subjects	Diabetic subjects	Hemodialysis subjects	OR1 (95%IC)	OR2 (95%IC)
-------------------	------------------	-------------------	-----------------------	-------------	-------------

246

247

248

249 **Table 2 : Paraoxonase 1 allelic frequency distribution in healthy, diabetic and**
250 **hemodialysis subjects**

251

252 *The distribution of alleles frequencies of each polymorphism was compared between case and*
253 *control subjects using χ^2 test. OR1 was calculated for diabetic versus healthy subjects,*
254 *whereas OR2 was calculated for hemodialysis versus healthy subjects.*

255

256 The gender-specific distribution of PON1 allelic frequencies in men showed Q allele
257 frequencies of $90.69 \pm 3.13\%$, $90.54 \pm 3.40\%$, and $81.54 \pm 3.40\%$ and R allele frequencies of
258 $9.30 \pm 3.13\%$, $9.46 \pm 3.40\%$, and $18.46 \pm 3.40\%$ in healthy, diabetic and hemodialysis
259 subjects, respectively (Table 3).

260 In women, the distribution of PON1 allelic frequencies showed Q allele frequencies of 83.72
261 $\pm 3.98\%$, $81.63 \pm 3.91\%$, and $75 \pm 6.24\%$, and R allele frequencies of $16.28 \pm 3.98\%$, $18.37 \pm$
262 3.91% , $25 \pm 6.24\%$ respectively in healthy, diabetic, and hemodialysis subjects, respectively
263 (Table 3).

264

265

Q	86.62±2.59%	85.46±2.69%	79.78±3%	1	1
R	13.37±2.59%	14.53±2.69%	20.22±3%	1.10(0.492.45)	1.64(0.773.49)

	Allelic frequency	Healthy subjects	Diabetic subjects	Hemodialysis subjects	OR1 (95%IC)	OR2 (95%IC)
Male	Q	90.69±3.13%	90.54±3.40%	81.54±3.40%	1	1
	R	9.30±3.13%	9.46±3.40%	18.46±3.40%	1.02(0.392.64)	2.21(0.955.13)
Female	Q	83.72±3.98%	81.63±3.91%	75±6.24%	1	1
	R	16.28±3.98%	18.37±3.91%	25±6.24%	1.16(0.562.41)	1.71(0.853.44)

266

267

268

269

270

271

272

273 **Table 3 : Paraoxonase 1 allelic frequency distribution by gender in healthy,**
274 **diabetic and hemodialysis subjects**

275

276 *The distribution of alleles frequencies of each polymorphism was compared between case and*
277 *control subjects using χ^2 test. OR1 was calculated for diabetic versus healthy subjects,*
278 *whereas OR2 was calculated for hemodialysis versus healthy subject.*

279

280 In men, the adjusted OR comparing the PON1 R allele with the Q allele was OR1 = 1.02
281 (95% CI: 0.392.64) in diabetic subjects and OR2 = 2.21 (95% CI: 0.955.13) in hemodialysis
282 subjects. In women the adjusted OR comparing the PON1 R allele with the Q allele was OR1
283 = 1.16 (95% CI: 0.562.41) in diabetic subjects and OR2= 1.71 (95% CI: 0.853.44) in
284 hemodialysis subjects (Table 3).

285 We also investigated the allelic frequency distribution of PON1 according to age. Among
 286 subjects aged 40-60 years, the frequencies of the PON1 Q allelic were $85.71 \pm 3.30\%$, 87.74
 287 $\pm 3.18\%$, and $74.44 \pm 4.60\%$, whereas the frequencies of the R allele were $14.29 \pm 3.30\%$,

Allelic frequency	Healthy subjects	Diabetic subjects	Hemodialysis subjects	OR1 (95%IC)	OR2 (95%IC)
-------------------	------------------	-------------------	-----------------------	-------------	-------------

288 $12.26 \pm 3.18\%$, $25.56 \pm 4.60\%$, in healthy, diabetic, and hemodialysis subjects, respectively
 289 (Table 4). In subject aged 40-60 years, the adjusted OR comparing the PON1 R allele with the
 290 Q allele was $OR1 = 0.84$ (95% CI: 0.371.90) for diabetic subjects and $OR2 = 2.06$ (95% CI: 1-
 291 4.22) for hemodialysis subjects.

292

293

294

295

296 **Table 4 : Paraoxonase 1 allelic frequency distribution as a function of age in**
 297 **healthy, diabetic and hemodialysis subjects**

298

299 *The distribution of allele frequencies of each polymorphism was compared between case and*
 300 *control subjects using χ^2 test. OR1 was calculated for diabetic versus healthy subjects.*
 301 *Whereas OR2 was calculated for hemodialysis versus healthy subjects.*

302 Among subject aged 60-80 years, the frequencies of the PON1 Q allele were $83.33 \pm 4.81\%$,
 303 $81.82 \pm 4.75\%$, and $85.22 \pm 3.78\%$, whereas the frequencies of the PON1 R allele were 16.67
 304 $\pm 4.81\%$, $18.18 \pm 4.75\%$, and $14.77 \pm 3.78\%$ in healthy, diabetic, and hemodialysis subjects,
 305 respectively (Table 4).

306 In subjects aged 60-80 years, the adjusted OR comparing the PON1 R allele with the Q allele
 307 was $OR1 = 1.11$ (95% CI: 0.532.31) for diabetic subjects and $OR2=0.87$ (95% CI: 0.41.86)
 308 for hemodialysis subjects.

[4060 years]	Q	85.71±3.30%	87.74±3.18%	74.44±4.60%	1	1
	R	14.29±3.30%	12.26±3.18%	25.56±4.60%	0.84(0.371.90)	2.06(14.22)
[6080 years]	Q	83.33±4.81%	81.82±4.75%	85.22±3.78%	1	1
	R	16.67±4.81%	18.18±4.75%	14.77±3.78%	1.11(0.532.31)	0.87(0.41.86)
	PON1			OR1 (95%IC)		OR2 (95%IC)
	Q			1		1
	QR			1.37 (0.62-3.04)		1.52 (0.69-3.35)

309 We also investigated the adjusted OR associated with the PON1 QQ, QR and RR phenotypes
310 according to age in diabetic and hemodialysis subjects. Among diabetic subjects aged 40-60
311 years, the adjusted OR comparing the QQ phenotype with the QR phenotype was OR1=1.15
312 (95% CI: 0.44-3.00), whereas the OR comparing the QQ phenotype with the RR phenotype
313 was OR1 = 0.35 (95% CI: 0.03-3.50). Among diabetic subjects aged 60-80 years, the adjusted
314 OR comparing the QQ phenotype with QR phenotype was OR1 = 2.08 (95% CI: 0.47-9.29),
315 whereas the OR comparing QQ phenotype with the RR phenotype was OR2=1.56 (95% CI:
316 0.24-10.19).

317

318

319

	RR	0.84 (0.22-3.25)	1.97 (0.63-6.21)
--	----	------------------	------------------

320 **Table 5 : Adjusted odd ratio in diabetic and hemodialysis subjects associated**
321 **with the risk of cardiovascular diseases**

	PON1	OR1 (95%IC)	OR2 (95%IC)
--	------	-------------	-------------

322

323

324

325 *OR1 was calculated for diabetic versus healthy subjects, whereas OR2 was calculated for*
326 *hemodialysis versus healthy subjects.*

327

328 Among hemodialysis subjects aged 40-60 years, the adjusted OR comparing the PON1 QQ
329 phenotype with the QR phenotype was OR2=1.33 (95% CI: 0.48-3.69), whereas the adjusted
330 OR comparing the QQ phenotype with the RR phenotype was OR2= 3.46 (95% CI: 0.83-
331 14.49) for RR. Among hemodialysis subjects aged 60-80 years, the adjusted OR comparing
332 the PON1 QQ phenotype with the QR phenotype was OR2 = 2.27 (95% CI: 0.56-9.27),
333 whereas the adjusted OR comparing QQ phenotype with the RR phenotype was OR2= 0.76
334 (95% CI: 0.10-5.76) (Table 5).

335

336

337

338

339

340

341 **Table 6 : Adjusted odd ratio in diabetic and hemodialysis subjects and the**
342 **associated risk of PON1 Q192R polymorphism**

343

	Q	1	1
[4060 years]	QR	1.15 (0.44-3.00)	1.33 (0.48-3.69)
	RR	0.35 (0.03-3.5)	3.46 (0.83-14.49)
[6080 years]	Q	1	1
	QR	2.08 (0.47-9.29)	2.27 (0.56-9.27)
	RR	1.56 (0.24-10.19)	0.76 (0.10-5.76)

344

345 *OR represents the adjusted odds ratio. OR1 was calculated for diabetic versus healthy*
 346 *subjects, whereas OR2 was calculated for hemodialysis versus healthy subjects.*

347

348 Discussion

349 Many studies support the role of oxidative stress in the pathogenesis of diabetes and chronic
 350 renal failure [16, 17, 18]. The main findings of this study were that, the PON1 Q192 R variant
 351 was more frequent in diabetic and hemodialysis subjects. Factors affecting serum PON1
 352 activity include type 1 and 2 diabetes mellitus, chronic renal disease, familial
 353 hypercholesterolemia, and inflammatory disorder such as rheumatoid arthritis and
 354 hemodialysis [19-22].

355 In our study, we observed an increase of Vit.E/TC ratio in PON1 QQ by comparison to RR ,
 356 indicating reduced lipid peroxidation and lower oxidative stress in subject with PON1 QQ. It
 357 is known that, the PON1 QQ inhibits the initiation and propagation of lipid peroxidation [23-
 358 25]. Vitamin E is a chain breaking antioxidant stopping the propagation of lipid peroxidation
 359 [26]. It also suggested that PON1 polymorphism may affect the ability of Vit.E to impede the
 360 development of atherosclerosis and to prevent inflammation.

361 We observed an increase of Vit E /CT ratio in PON1 QQ carriers by comparison to PON1 RR
 362 carriers in diabetics and hemodialysis indicating changes in lipid profile in these subjects.
 363 Aviram et al. reported that, PON1 inhibits efficiently the production of lipid peroxides at 61
 364 % and aldehyde compounds at 58 % [23-25]. PON1 also inhibits LDL oxidation efficiently if
 365 it presents in initiation of lipid peroxidation step. But the PON1 RR inhibits production of
 366 lipid peroxidation at 46 % and aldehyde compounds in 38% [23-25].

367 Published data on the association between PON1 polymorphisms and coronary heart disease
368 have yielded controversial results [27]. Study from Bub et al. indicates that, PON 1 Q192R
369 polymorphism is associated with reduced lipid peroxidation in R-allele-carriers but not in QQ
370 homozygous elderly subjects on a tomato-rich diet [27].

371 The relationship between the two human PON1 amino acid variants, the Leu55Met and the
372 Gln192Arg polymorphism, and the risk of cardiovascular disease is also documented in this
373 study. The main purpose is to investigate the link between the PON1 allele frequency
374 distribution, adjusted odds ratio and the risk of cardiovascular diseases development in a
375 Moroccan population.

376 Our result showed a high risk of cardiovascular disease in QR carriers vs QQ carriers in
377 diabetic [OR1= 1.37; 95% CI: 0.62-3.04] and hemodialysis subjects [OR2=1.52; 95% CI:
378 0.69-3.35].

379 The risk of cardiovascular diseases in QR and RR diabetic and hemodialysis subjects is due to
380 Gln192Arg polymorphism of PON1. The Gln192Arg polymorphism of PON1 indicates
381 changes from glutamine (the Q variant) to arginine (the R variant) at position 192 of the
382 amino acid sequence [8,9]. These changes may affect its hydrolytic activity [23, 24]. Arginine
383 is a conditionally essential amino acid serving as a substrate for protein synthesis, L-arginine
384 is the pre- cursor for nitric oxide (NO), is responsible for free radical production [[9, 28].
385 These results are in line with other studies that investigated PON1 polymorphism related to
386 cardiovascular diseases risk [19, 29, 30].

387 We also observed that, the PON1 192Q allele frequency (86.62%; 85.46% and 79.78%) is
388 higher than that of PON1 192R allele frequency (13.37%; 14.53% and 20.22%) respectively
389 in healthy, diabetic and hemodialysis subjects. Van Den Berg et al. observes comparable R-
390 allele frequencies for subjects with normal Glucose Tolerance 32% and Newly diagnosed
391 Type 2 diabetes 25% [31]. This study confirms that, the PON1 192 R allele is responsible for
392 cardiovascular disease in diabetic and hemodialysis patients.

393
394 In the present study, we have shown that, PON1 192 R allele provides lowest protection
395 against oxidative stress in diabetic and hemodialysis patients. As a function of age, we note a
396 high risk of cardiovascular diseases in hemodialysis [OR2=2.06; 95% CI: 1-4.22] and diabetic
397 subjects [OR1=1.11; 95% CI: 0.53-2.21] when comparing allelic frequencies homozygosity
398 for QQ variant and RR variant. These results suggest that, oxidative stress damage is higher in
399 our subjects.

400
401 Paraoxonase 1 Arg 192 Gln allelic frequency distribution showed a high frequency of the R
402 allele in female diabetic. Our diabetic women were obese and hypertensive. Hypertension

403 affects the PON1 activity and also increases the susceptibility to atherosclerosis [32]. It also
404 compromises the capacity of their HDL to prevent the accumulation of lipid peroxide on
405 human LDL [33]. Lawlor et al. study indicated that, there was a high risk of cardiovascular
406 diseases among the findings from the British Women's Heart and Health cohort study and a
407 meta-analysis over 60 years [34]. PON1 allelic frequency distribution is elevated in male and
408 female hemodialysis patients as a result of dialysis process. Dialysis process produces free
409 radicals responsible for oxidative damage in these subjects [35, 36]. Hemodialysis alters also
410 lipid profile, total antioxidant capacity, vitamin A, E and C concentration in humans and also
411 increase cardiovascular diseases development [37]. To our knowledge, few studies provide
412 data of the PON1 Q192R polymorphism related to cardiovascular diseases in hemodialysis
413 patients.

414 Our study has shown that, diabetic subjects and hemodialysis patients carrying RR and QR
415 had high triglycerides level when compared to healthy control carrying QQ. These results are
416 in line with others studies [38, 39]. Jarvik et al. reported that PON1 is a better predictor of
417 vascular diseases than be the PON1-192 or PON1-55 [38]. This study showed that the R allele
418 alters the reactivity of the paraoxonase [38]. Our result which is in agreement with other
419 studies suggested that PON1 activity can be used as a cardiovascular disease prediction
420 marker in healthy, diabetic and hemodialysis patients [38, 39]. Our study confirms that, PON1
421 polymorphisms affects the lipid profile, particularly it showed that the R allelic frequency of
422 PON1 disturbs serum triglycerides distribution in lipoproteins. Compared with hemodialysis
423 patients, diabetic subjects had better lipid profile (lower triglyceride) because they reported a
424 higher use of lipid medication. Deakin et al. reported that Statins modulates the expression of
425 the PON1 gene and increase serum Paraoxonase [40]. A logical explanation appears to be the
426 higher frequency of the R allele in our hemodialysis patients. These findings point at less
427 protection against oxidation and higher risk of cardiovascular diseases at the beginning of
428 chronic renal disease [40, 41]. A plausible explanation can be the high level of triglycerides
429 in Hemodialysis subjects which alter the composition of HDL, VLDL, LDL cholesterol, and
430 finally influence the PON1 binding and its activity [41].

431
432 In our study, the low risk of CVD observed in Morocco is due to Mediterranean diet.
433 Mediterranean diet generically describes an eating pattern. In Morocco, the diet is rich in
434 argan oil, fruit and vegetable. The benefits of this diet is its content in antioxidant particularly
435 Vitamin E. The Mediterranean diet lowers the risk of heart disease and early death.

436 In conclusion, this study investigates the relationship between PON1 phenotype distribution
437 and the risk of cardiovascular disease development in healthy, diabetic and hemodialysis
438 patients. Our findings suggest a slight increase in cardiovascular risk, particularly among
439 carriers of the R allele.

440

441 **References**

442

443 [1] Foley R.N., Parfrey P.S. and Sarnak M.J.. Epidemiology of cardiovascular diseases in
444 chronic renal disease. *J Am Soc Nephrol* 9 (1998) 16-23.

445 [2] Gbandjaba N.Y., Hassar M., Saïle R., Berrougui H., Taki H., Lebrazi H., Ghalim N.,
446 Khalil A. Paraoxonase activity in healthy, diabetic and hemodialysis patients. *Clin Biochem.*
447 45 (2012) 470-474.

448 [3] Murray J.L., Lopez A.D . Global mortality, disability, and the contribution of risk factors :
449 Global Burden of Disease Study. *Lancet* 349 (1997) 1436-1442.

450 [4] Ross R. The pathogenesis of atherosclerosis : a perpective for the 1990s. *Nature* 362
451 (1993) 801-809

452 [5] Lusis A.J. Atherosclerosis. *Nature* 407 (2000) 233-241.

453 [6] Mackness B., Durrington P.N., Mackness M.I. Human serum paraoxonase. *Gen*
454 *Pharmacol.* 31 (1998) 329-336.

455 [7] Watson A.D., Berliner J.A., Hama S.Y., La Du B.N., Faull K.F., Fogelman A.M., Navab
456 M. Protective effect of high density lipoprotein associated paraoxonase. Inhibition of the
457 biological activity of minimally oxidized low density lipoprotein. *J Clin Invest* 96 (1995)
458 2882-2891.

459 [8] Smolen A., Eckerson H.W., Gan K.N., Hailat N., La Du B.N. Characteristics of the
460 genetically determined allozymic forms of human serum paraoxonase/arylesterase. *Drug*
461 *Metab Dispos* 19 (1991) 107-112.

462 [9] Mogarekar M.R., Chawhan S.S. The determination of Q192R polymorphism of
463 paraoxonase 1 by using non toxic substrate p-nitrophenylacetate. *Indian J Hum Genet* 19
464 (2013) 71-77

465 [10] Mahrooz A., Alizadeh A., Gohari G. The salt stimulation property of serum paraoxonase
466 (PON1) could be a valuable factor in evaluating the enzyme status in ischemic stroke : the
467 role of activity-determined PON1 192Q/R phenotypes. *J Neurol Sci* 338 (2014) 197-2002.

468 [11] Costa L.G., Li W.F., Richter R.J., Shih D.M., Lusis A., Furlong C.E.. The role of
469 paraoxonase (PON1) in the detoxication of organophosphates and its human polymorphism.
470 *Chem Biol Interact* 119-120 (1999) 429-438.

471 [12] Lahrach H., Essiarab F., Timinouni M., Hatim B., El Khayat S., Er-Rachdi L., Jarir J.,
472 Kettani A., Ghalim N., Taki H., Lebrazi H., Ramdani B., Saïle R.. Association of
473 apolipoprotein E gene polymorphism with end-stage renal disease and hyperlipidemia in
474 patients on long-term hemodialysis. *Ren Fail.* 26 (2014) 1-6.

- 475 [13] Ferretti G., Bacchetti T., Busni D., Rabini R.A., Curatola G. Protective effect of
476 paraoxonase activity in high-density lipoproteins against erythrocyte membranes peroxidation
477 : a comparison between healthy subjects and type 1 diabetic patients. *J Clin Endocrinol*
478 *Metab.* 89 (2004) 2957-2962.
- 479 [14] Mackness B., Durrington P.N., Mackness M.I. Human serum paraoxonase. *Gen*
480 *Pharmacol.* 31 (1998) 329-336.
- 481 [15] Deakin S.P., James R.W. Genetic and environmental factors modulating serum
482 concentrations and activities of the antioxidant enzyme paraoxonase-1. *Clin Sci.* 107 (2004)
483 435-447.
- 484 [16] Cerit N., Onuk A. A., Ellidag H.Y., Eren E., Bulbuler N., Yilmaz N. Arylesterase and
485 oxidative stress in operating room personnel. *Adv Clin Exp Med.* 23 (2014) 49-55
- 486 [17] Bozkürk M.B, Günay E., Ogan N., Candemir İ., Eker U., Öztürk A. Serum paraoxonase-
487 1 as a marker of oxidative stress and pulmonary dysfunction in sarcoidosis: association with
488 disease activity and prognostic potential. *Front Immunol.* 16 (2026) 1731991. doi:
489 10.3389/fimmu.2025.1731991. eCollection 2025.
- 490 [18] Gašperšič R., Taler-Verčič A., Petrič B., Goličnik M., Bavec A. Total esterase and
491 paraoxonase activity in human saliva of periodontitis patients and healthy individuals; a pilot
492 study. *Chem Biol Interact.* 423 (2026) :111836. doi: 10.1016/j.cbi.2025.111836.
- 493 [19] Delibaş E.A.Ö, Köse K., Yazici C., Tokgöz B. Oxidative Stress-Related HDL
494 Dysfunction in Hemodialysis: The Clinical Utility of MPO/PON1 and MPO/HDL-C Ratios in
495 Cardiovascular Risk Assessment. *Ther Apher Dial.* 30 (2026) 78-84. doi: 10.1111/1744-
496 9987.70094. Epub 2025 Nov 9.
- 497 [20] Parfentyeva E., Saha S., Hjellset V. T., Kopprasch S., Schwarz P. E. Assessment of small
498 C-Fiber status for screening of oxidative stress in patients at high risk of Diabetes. *Horm*
499 *Metab Res* 46 (2014) 360-364
- 500 [21] Bernal-Hernandez Y.Y., Medina-Diaz I.M., Barron-Vivanco B.S., Robledo-Marengo
501 MdeL, Giron-Perez M.I., Perez-Herrera N.E., Quintanilla-Vega B., Cerda-Florez R., Rojas-
502 Garcia A.E. Paraoxonase 1 and its relationship with pesticide biomarkers indigenous mexican
503 farmworkers. *J Occup Environ Med.* 56 (2014) 291-290.
- 504 [22] Paragh G., Törocsik D., Seres I, Harangi M., Illyés L., Balogh Z., Kovács P. Effect of
505 short term treatment with simvastatin and atorvastatin on lipid and paraoxonase activity in
506 patients with hyperlipoproteinaemia. *Curr Med Res Opin.* 20 (2004) 1321-1327.
- 507 [23] Aviram M., Rosenblat M., Bisgaier C.L., Newton R.S., Primo-Parmo S.L., La Du. B.N.
508 Paraoxonase inhibits high-density lipoprotein oxidation and preserves its functions. A
509 possible peroxidative role for paraoxonase. *European journal of clinical investigation.* (1998a)
510 1581-1590.
- 511 [24] Aviram M., Rosenblat M., Billecke S., Eurogul J., . Sorenson R, Bisgaier C.L., Newton
512 R.S., La Du B.N. Human serum paraoxonase (PON1) is inactivated by oxidized low density

- 513 lipoprotein and preserved by antioxidants. *Free Radical Biology and Medicine*. 26 (1999)
514 892-904
- 515 [25] Aviram M., Billecke S., Sorenson R., Bisgaier C., Newton R., Rosenblat M., Eroglu J.,
516 Hsu C., Dunlop C., La Du B. Paraoxonase active site required for protection against LDL
517 oxidation involves its free sulfhydryl group and is different from that required for its
518 arylesterase/paraoxonase activities : selective action of human paraoxonase allozymes Q and
519 R. *Arterioscler Thromb Vasc Biol* 18 (1998b) 1617-1624.
- 520 [26] Halliwell B. The chemistry of free radicals. *Toxicology and industrial health*. 9 (1993) 1-
521 21.
- 522 [27] Bub A., Barth S., Watzl B., Briviba K., Herbert B.M., Lührmann P.M., Neuhäuser-
523 Berthold M., Reckemmer G. Paraoxonase 1 Q192R (PON1-192) polymorphism is
524 associated with reduced lipid peroxidation in R-allele-carrier but not in QQ homozygous
525 elderly subjects on a tomato-rich diet. *J Nutr*. 134 (2004) 1081-3.
- 526 [28] Diepeveen S.H., Verhoeven G.H., Van Der Palen J., Dikkeschei B.L., Van Tits L.J.,
527 Kolsters G., Offerman J.J., Bilo H.J., Stalenhoef A.F. Oxidative stress in patients with end-
528 stage renal disease prior to the start of renal replacement therapy. *Nephron Clin Pract* 98
529 (2004) 3-7.
- 530 [29] Watzinger N., Schmidt H., Schumacher M., Schmidt R., Eber B., Fruhwald F.M.,
531 Zweiker R., Kostner G.M., Klein W.. Human Paraoxonase1 Gene Polymorphisms and the
532 Risk of Coronary Heart Disease: A Community-Based Study. *Cardiology* (2002) 98 (3): 116–
533 122.
- 534 [30] Newton R.S., La Du B.N. Human serum paraoxonase (PON1) is inactivated by oxidized
535 low density lipoprotein and preserved by antioxidants. *Free Radical Biology and Medicine*.
536 26 (1999) 892-904.
- 537 [31] Van Den Berg S.W., Jansen E.H., Kruijshoop M., Beekhof P.K., Blaak E., Van Der
538 Kallen C.J., Van Greevenbroek M.M., Feskens E. Paraoxonase 1 phenotype distribution and
539 activity differs in subjects with newly diagnosed Type 2 diabetes (the CODAM Study).
540 *Diabetic medicine*. 25 (2008) 186-193.
- 541 [32] Diyane K., El Ansari N., El Mghari G., Anzid K., Cherkaoui M. Characteristics of the
542 association type 2 diabetes and hypertension in the elderly aged 65 and over. *Pan Afr Med J*.
543 14 (2013) 100.
- 544 [33] Reaven P. D., Napoli C., Merat S., Witztum J.L. Lipoprotein modification and
545 atherosclerosis in aging. *Exp Gerontol* 34 (2000) 527-537.
- 546 [34] Lawlor D. A., Day I. N. M. , Gaunt T. R., Hinks L. J, Timpson N., Ebrahim S., Smith G.
547 D.. The association of the paraoxonase (PON1) Q192R polymorphism with depression in
548 older women: findings from the British Women’s Heart and Health Study. *Journal of*
549 *epidemiology and community health*. 61 (2007) 85-7.

- 550 [35] Daschner M., Lenhartz H., Botticher D. Influence of dialysis on plasma lipid
551 peroxidation products and antioxidant levels. *Kidney Int* 50 (1996) 1262-1268.
- 552 [36] Lacson E. Jr. and Lazarus J.M. Dialyzer best practice : single use or reuse ?. *Semin Dial*
553 19 (2006) 120-128.
- 554 [37] El Muhtaseb M.S., Talwar D., Duncan A., O'reilly D.S., McKee R.F., Anderson J.H.,
555 Foulis A., Finlay I.G. Free radical activity and lipid soluble anti-oxidant vitamin status in
556 patients with long-term ileal pouch-anal anastomosis. *Colorectal Dis* 11 (2008) 67-72.
- 557 [38] Jarvik J.P., Rosek L.S., Brophy B.H., Hatsukami T.S., Richter R.J., Schellenberg G.D.,
558 Furlong C.E. Paraoxonase (PON1) phenotype is a better predictor of vascular disease than is
559 PON1 (192) or PON1(55) genotype. *Atherosclerosis, thrombosis, and vascular biology*. 20
560 (2000) 2441-2447
- 561 [39] Gugliucci A. Activation of paraoxonase 1 is associated with HDL remodelling ex vivo.
562 *Clin Chem Acta*. 429 (2014) 38-45
- 563 [40] Deaking S., Leview I., Brulhurt-Meynet MC, James RW. Paraoxonase 1 promoter and
564 serum paraoxonase : a predominante role for polymorphic-107 implicating the Sp1 factor.
565 *Biochem J* 372 (2003) 643-649
- 566 [41] Senti M., Tomás M., Marrugat J., Elosua R. Paraoxonase1-192 polymorphism
567 modulates the nonfatal myocardial infarction risk associated with decreased HDLs.
568 *Atherosclerosis, Thrombosis and Vascular biology*. 21 (2001) 415-420.

569

570

571

572

573