

1 **In Silico Analysis of Differentially Expressed Gene Sets Reveals LY6E as a Potential**
2 **Candidate Gene in Oral Squamous Cell Carcinoma (OSCC) Progression.**

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6
7 **Abstract**

8 Oral Squamous Cell Carcinoma (OSCC) is most prevalent head and neck cancer and potentially
9 malignant in oral cavity cancer. OSCC is asymptomatic in the early stages and mostly
10 diagnosed at late-stage that significantly hinders successful treatment. Genetic mutations may
11 also cause cancer development in oral cavity; however, no specific genes or markers has been
12 identified for OSCCs progression. While a few studies have reported some molecular markers
13 like Cyclin D1 and BCL2, the lack of specific early diagnostic and prognostic markers remains
14 the biggest obstacle for OSCC therapy till date. This study has been designed to identify
15 specific molecular markers for precise diagnosis and prognosis of OSCC for successful
16 treatment. Understanding the complex molecular mechanisms involved in OSCC pathogenesis
17 is crucial to address this challenge effectively. The study utilizes a comprehensive approach to
18 examine the differentially expressed genes (DEGs) in a large OSCC dataset (GSE85195)
19 obtained from the Gene Expression Omnibus (GEO) database. Upon analysis of datasets, we
20 found 245 common DEGs in all four stages of cancer within the dataset. Further network
21 analysis of 245 DEGs revealed that 10 genes are highly connected based on the highest
22 MCODE (Molecular Complex Detection) score. Among all 10 genes, LY6E gene showed
23 highest MCODE value, indicating LY6E may be critically involve in OSCC progression and
24 act as promising candidate biomarker. To delineate the molecular mechanism of gene
25 regulation of LY6E, we further looked into its interactions with microRNAs (miRNAs) and
26 transcription factors (TFs). Our findings identified certain miRNAs and TFs that may regulate
27 LY6E expression, indicates its involvement in OSCC development. This study emphasizes the
28 potential role of LY6E in OSCC progression and suggests LY6E could serve as a valuable
29 diagnostic tool. These findings lay the groundwork for future research to develop targeted
30 therapies based on the LY6E signaling network, which could ultimately improve clinical
31 outcomes for OSCC patients.

33 **Keywords:** Oral Squamous Cell Carcinoma (OSCC), Differentially Expressed Genes (DEGs),
34 Extracellular Matrix (ECM) Receptor Interaction, Matrix Metalloproteinases (MMPs),
35 Signaling Pathways (e.g., IL-17, Wnt, CCKR)

37 **Introduction**

38 Oral Squamous Cell Carcinoma (OSCC) represents the leading cancer type in the oral cavity,
39 comprising more than 90% of the cases [1]. It occurs due to the uncontrolled growth of altered
40 squamous epithelial cells that line the oral cavity. It is the sixth most common cancer in the
41 world [2, 3]. The majority of oral cancer cases are found in South Asia, primarily due to the
42 prevalent habits of smoking, alcohol consumption, and betel quid chewing in the region
43 [4]. Oral cancer is 2 to 3 times more frequent in men than in women. In India, it ranks second in
44 terms of incidence, after breast cancer, and third in terms of mortality, following breast and
45 cervix uteri cancer. Cancers originating in the lip and oral cavity represent major contributors to
46 cancer-related morbidity and mortality, accounting for an incidence rate of 10.3% and a
47 mortality rate of 8.8%, according to Globocan 2020 data (<https://gco.iarc.fr>). The primary
48 causative factors for the development of oral cancer are tobacco and alcohol consumption,
49 estimated to be responsible for 90% of cases [3, 5]. Additionally, the Human Papillomavirus
50 (HPV) and Epstein-Barr virus (EBV) are also crucial biological risk factors for oral cancer
51 development [6].

52 The development of OSCC is a multistep process modulated by endogenous as well as
53 environmental factors and involves changes in normal mucosa accompanied by invasion and
54 distant proliferation to lymph node [7][8]. During the development of oral cancer, multiple
55 genetic and epigenetic events occur that alter the normal functions of oncogenes and tumor
56 suppressor genes, resulting in the progression [9][7]. Interferon beta (IFN- β) is critical in the
57 body's defense against diseases and impacts cancer progression and viral infections [10]. It
58 regulates the immune system and inhibits viral replication, thereby reducing the risk of virus-
59 associated carcinogenesis [11]. The LY6E gene, which is involved in immune response
60 regulation, has been identified as a significant marker in Oral Squamous Cell Carcinoma
61 (OSCC) research, suggesting a potential role in cancer progression through immune evasion
62 mechanisms [12]. IFN- β 's interaction with LY6E could be pivotal, especially since LY6E
63 influences the tumor's ability to evade the immune system [13]. This relationship is further
64 complicated by the role of viruses in cancer development, where LY6E's modulation of the
65 immune response against viruses could intersect with cancer therapy, particularly in therapies
66 aiming to bolster immune surveillance [14].

67 Detecting oral cancer in its early stages and potential treatments is considered the most
68 efficient way to improve patient survival. The 5-year survival chance is less than 50% in late-
69 stage diagnosis. As a general rule, the prognosis of disease worsens with the advancement
70 [15][16]. However, if diagnosed in an early stage, the survival rates can exceed 80%[16].
71 Regardless of all favorable therapeutic advancements in the field of oral cancer, there are no
72 targeted curative treatments, and overall survival remains at a disappointingly stable
73 level[17]. Several molecular markers implicated in the carcinogenesis of OSCC have been
74 evaluated by many investigators, including molecules involved in cell cycle regulation,
75 apoptosis, angiogenesis, DNA repair system, and degradation of extracellular matrix.
76 However, this evidence remains inconclusive; to battle the poor morbidity and mortality rate
77 associated with OSCC, there is a great need to evaluate more effective targeted treatment
78 options[18][19]. Currently, the main conventional prognostic factors for the survival of oral
79 squamous cell carcinoma patients are the histological identification of tumor at the time of
80 diagnosis; however, recent studies use many prognostic biomarkers from the body fluids of
81 oral cancer patients which are known to influence the oncological outcomes [20][21].
82 Nowadays, cancer research is focused on understanding the underlying mechanisms, which can
83 be crucial to finding novel pharmacological markers for the treatment of OSCC. Yet not all
84 aspects of the progression of tumor have been fully understood[22][23]. Moreover, identifying
85 novel molecular markers for early detection, prognosis stratification, and therapeutic
86 evaluation is a continuous challenge[24]. However, various studies on markers are available.
87 Still, the numerous molecular genes are tightly connected in migration, proliferation, apoptosis,
88 and metastasis, which makes it challenging to understand the OSCC mechanism and detection
89 at the early stage [23]. Identifying improved targeted treatment and prognostic markers are
90 significant priorities in oral cancer. It requires knowledge of oral cancer's genetics and
91 molecular biology [25].
92 Therefore, with this study, we hope to shed light on promising biomarkers and their role in
93 OSCC progression that has not been explored yet. There is a significant clinical need to build a
94 model for developing novel targeted approaches against oral cancer. This study analyzed the
95 oral cancer dataset GSE85195, which had 34 OSCC, 15 pre-cancer, and 1 control sample. A
96 total of 245 genes were identified, which is common DEGs. Further analysis revealed that 10
97 genes are highly connected based on the highest MCODE score, indicating that LY6E is a
98 promising molecular driver for oral cancer. The study highlighting LY6E as a candidate
99 biomarker for oral cancer underscores the need to understand how LY6E's regulation of
100 immune responses could be leveraged in therapeutic contexts, possibly enhancing

101 immunotherapy's effectiveness. The connection between IFN- β , LY6E, and cancer progression
102 underscores the potential for new therapeutic approaches by targeting these mechanisms,
103 especially in cancers with viral involvement or where immune evasion is a principal concern.
104 This approach may lead to the development of target-dependent tumor therapy.

105

106 **Materials and Methods**

107 **Dataset Analysis**

108 Gene expression profiles for 50 oral cavity samples, including 34 cases of oral squamous cell
109 carcinoma (OSCC), 15 cases of oral leukoplakia (OLK), and 1 control sample were collected
110 from the NCBI-GEO database, available at (<http://www.ncbi.nlm.nih.gov/geo/>). These profiles
111 were gathered using the GSE accession ID GSE85195 and are based on the GPL6480 platform.

112

113 **Identification of DEGs**

114 The GEO2R (<https://www.ncbi.nlm.nih.gov/geo/geo2r/>) tool was utilized to categorize
115 samples into relevant groups (Control, Stage 1, Stage 2, Stage 3, and Stage 4) and to facilitate
116 comparisons between these groups. The analysis of differential expression was conducted
117 using the GEOquery and Limma tools. GEOquery serves to parse GEO data into R data
118 structures, while limma (Linear Models for Microarray Analysis) acts as a statistical test
119 designed to pinpoint differentially expressed genes in microarray data. The sorting of
120 differentially expressed genes (DEGs) was based on the LogFC10 value (+/- >1) and the
121 adjusted p-value. DEGs among the different groups (Control, Stage 1, Stage 2, Stage 3, and
122 Stage 4) were identified, and those common across all groups, including the control, were
123 compared to uncover common gene signatures.

124

125 **Network analysis and module extraction**

126 Identified gene sets were fed to the STRING (<https://string-db.org/>), and the protein-protein
127 interactions (PPI) were visualized among them using Cytoscape v 3.8.2
128 (<https://cytoscape.org/>). Gene pairs with a combined score greater than 0.4 were considered for
129 networking. The top modules were extracted using the MCODE (Molecular Complex
130 Detection) plugin of Cytoscape. Parameters taken for the module extraction were degree
131 cutoff= 2, Node score cutoff = 0.2, K-score = 2, and Max. depth=100.

132

133

134

135 **Gene ontology and KEGG pathway enrichment analysis of DEGs**

136 The significant enrichment analysis of DEGs was assessed in this study based on Gene
137 ontology (GO) using gProfiler (<https://biit.cs.ut.ee/gprofiler/gost>), a biological database of
138 gene/protein families and a free online tool for functional annotation analysis. GO
139 analysis includes the following categories: biological process (BP), cellular component (CC),
140 molecular function (MF), and panther pathway (PP). The gProfiler is a common and useful
141 method for annotating genes and gene products and identifying characteristic biological
142 attributes of high-throughput genome or transcriptome data. KEGG pathway enrichment
143 analysis was performed using the gProfiler platform in the biological context of differentially
144 expressed genes.

145

146 **Expression profile, stage plot, and survival analysis**

147 The expression of identified biomarkers was further validated with TCGA and GTEx datasets
148 of Liver hepatocellular carcinoma in 369 tumors and 160 normal cases using the GEPIA
149 (<http://gepia.cancer-pku.cn>) tool. Log₂FC cut-off and p-value cut-off were taken as 1 and 0.01,
150 respectively. Tumor samples were indicated in red while normal samples were indicated in
151 grey. Survival analysis was also performed using the GEPIA tool. Overall survival with a
152 median group cut-off at a 95% confidence interval and a cut-off high as low as 50% was taken
153 as criteria. Overall survival analysis for each gene selected based on the MCODE score was
154 performed.

155

156 **Transcription factor prediction analysis**

157 Transcription factors and biomarkers-related micro-RNAs have been identified using the
158 Network analyst tool (<https://www.networkanalyst.ca/>). Networkanalyst is used for
159 gene/protein list. The gene list is pasted in the gene list input tab; *Homo sapiens* were selected
160 and uploaded for analysis.

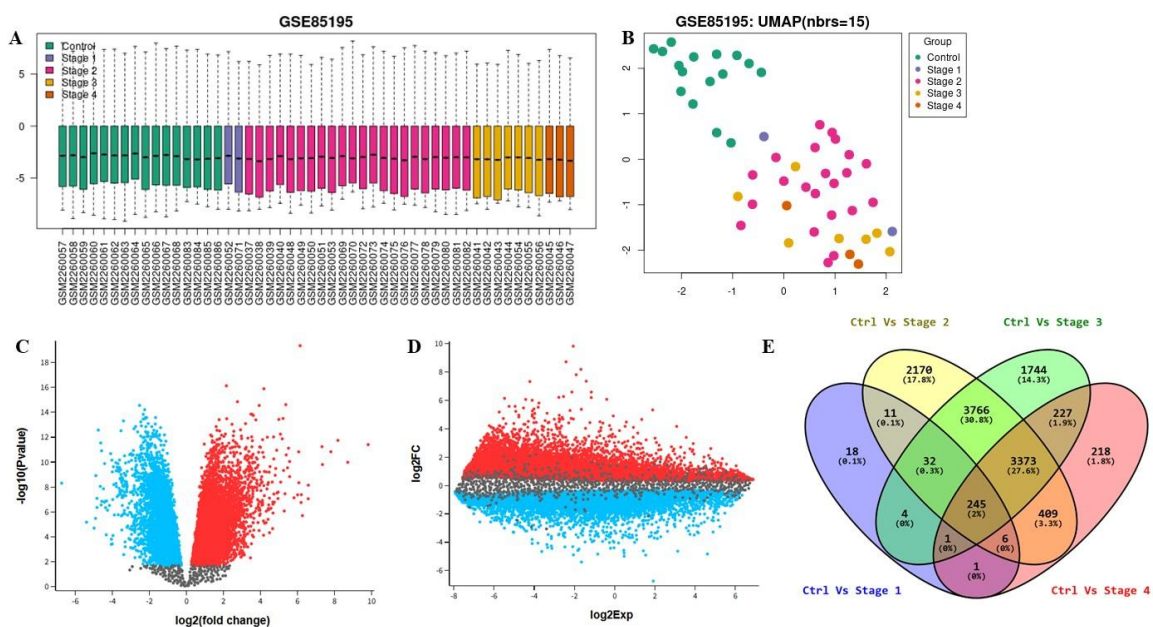
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162 **Results**

163 **Uncovering 245 key DEGs in oral squamous cell carcinoma (OSCC) progression.**

164 The GSE Id GSE85195 expression profile is analyzed using GEO2R with the GEO-query and
165 limma package, grouping samples into Control, Stage 1, Stage 2, Stage 3, and Stage 4. The box
166 plot (Fig. 1A) demonstrates the normalization of sample data within the dataset, offering a
167 clear view of distribution and central tendencies. The UMAP plot in (Fig. 1B) provides a
168 detailed visualization of relationships between different sample groups, including those that

169 appear distantly connected. The volcano plot (Fig. 1C) and the MD-Plot (Fig. 1D) reveal
 170 substantial evidence of differential gene expression across all sample groups, highlighting
 171 significant genetic variations and patterns. Our analysis identified sets of common DEGs in the
 172 dataset GSE85195 based on both magnitude of change (\log_2 fold change ± 1) and statistical
 173 significance (adjusted p-value < 0.05) in all comparisons: Control vs. Stage 1 (318 DEGs),
 174 Control vs. Stage 2 (10012 DEGs), Control vs. Stage 3 (9392 DEGs), Control vs. Stage 4
 175 (4480 DEGs). Further comparison employing Venn diagrams revealed 245 significantly
 176 differentially expressed genes (DEGs), which will serve as early biomarkers of OSCC as these
 177 are present in the control, early stage as well and advanced stages of OSCC (Fig. 1E).

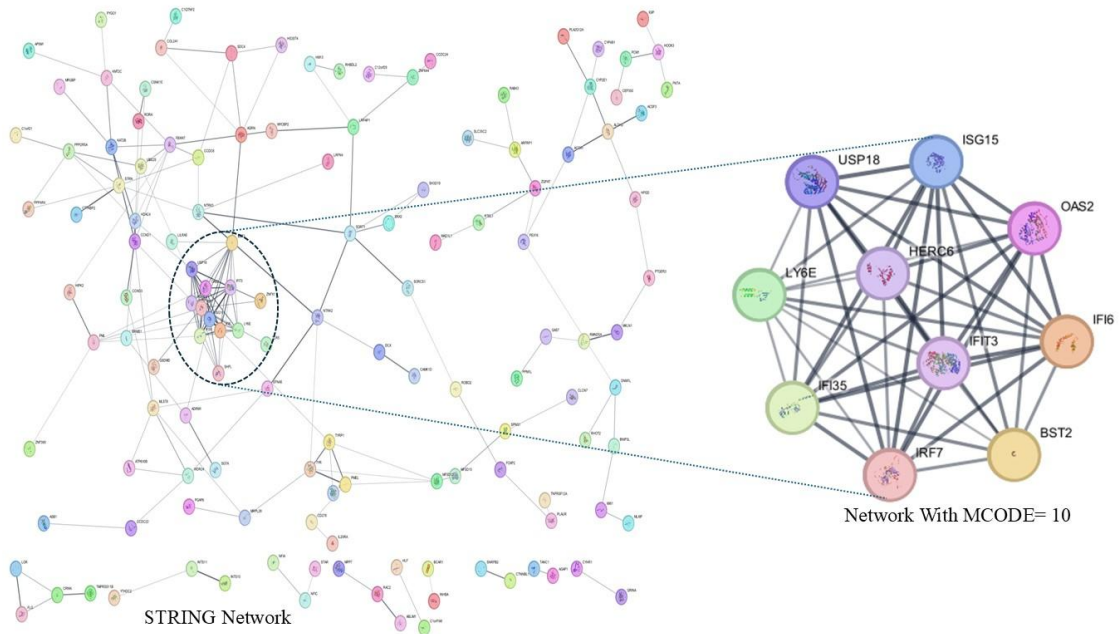


178
 179 **Figure. 1: Expression Analysis of DEGs:** A- Data Normalization Box Plot, B- UMAP for
 180 Group Expression, C- Volcano Plot, D- MD Plot, E- Venn Diagram For Common DEGs in all
 181 stages of Oral Cancer.

182 Construction of network, analysis, and extraction of modules

183 All 245 DEGs were then subjected to STRING analysis (<https://string-db.org/>) and network
 184 analysis by Cytoscape. STRING analysis resulted in a protein-protein interaction network with
 185 a combined score > 0.4 . The network was visualized further in Cytoscape, and module
 186 extraction was performed using the MCODE plugin. Our data analysis revealed a highly
 187 interconnected module of 10 DEGs with an MCODE score of 10, suggesting a potential
 188 candidate for OSCC progression. These 10 DEGs include USP18 (Ubiquitin specific peptidase
 189 18), OAS2 (2'-5'-oligoadenylate synthetase 2), LY6E (Lymphocyte antigen 6 family member
 190 E), ISG15 (Interferon-stimulated gene 15), IRF7 (Interferon regulatory factor 7), IFIT3

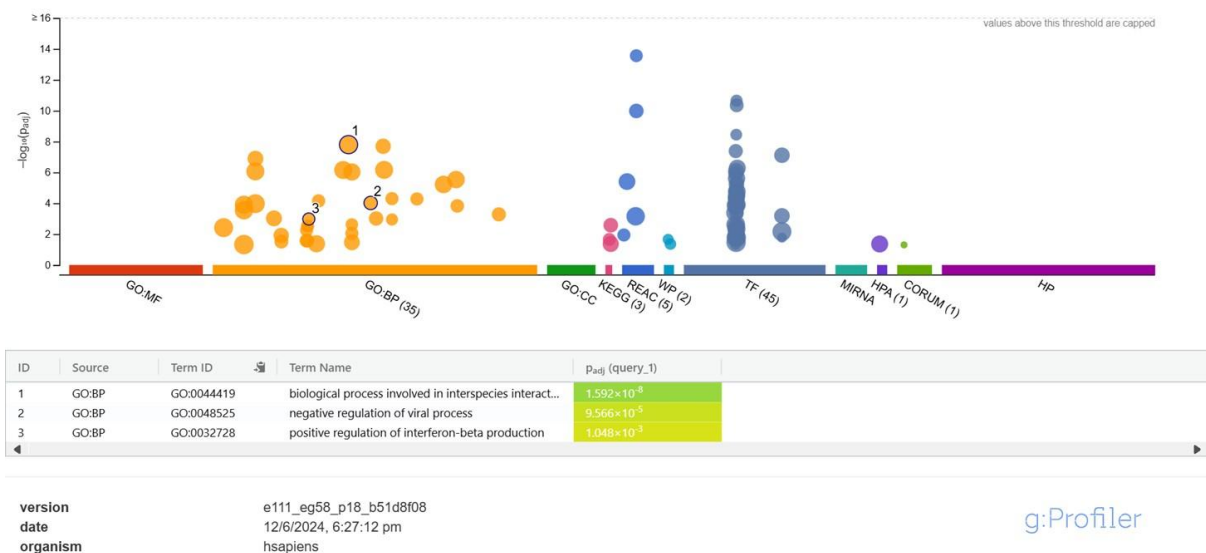
191 (Interferon Induced Protein With Tetratricopeptide Repeats 3), IFI6 (Interferon-alpha inducible
 192 protein 6), IFI35 (interferon-induced protein 35), HERC6 (HECT And RLD Domain
 193 Containing E3 Ubiquitin Protein Ligase Family Member 6), BST2 (Bone marrow stromal
 194 antigen 2) (Fig 2). Subsequently, the 10 genes within this module with the highest individual
 195 MCODE scores were selected for further pathway analysis.



196
 197 **Figure. 2: Analysis and Extraction of Modules with Highest MCODE Score.** Network
 198 visualization from STRING analysis showing modules with the highest MCODE score of 10.
 199 Nodes are color-coded to highlight these high-score modules, emphasizing their dense
 200 interconnections and central role in the network.

201 **Gene ontology (GO) analysis of key DEGs in OSCC**

202 Unveiling the functional underpinnings of OSCC progression, Gene Ontology analysis of these
 203 10 high-scoring STRING modules (MCODE score 10) was performed. GO analysis performed
 204 by gProfiler tools revealed a remarkable enrichment for processes critical for cancer cell
 205 proliferation and cell motility (Fig. 3, Table 1). These genes, intimately linked to the biological
 206 process involved in interspecies interaction between organisms, negative regulation of the viral
 207 process, and positive regulation of interferon-beta production, seem to likely play a pivotal role
 208 in orchestrating the uncontrolled growth and invasive behavior of cancerous cells. This
 209 convergence of key regulatory functions in a single module highlights its potential as a
 210 promising therapeutic target for combating this aggressive disease.

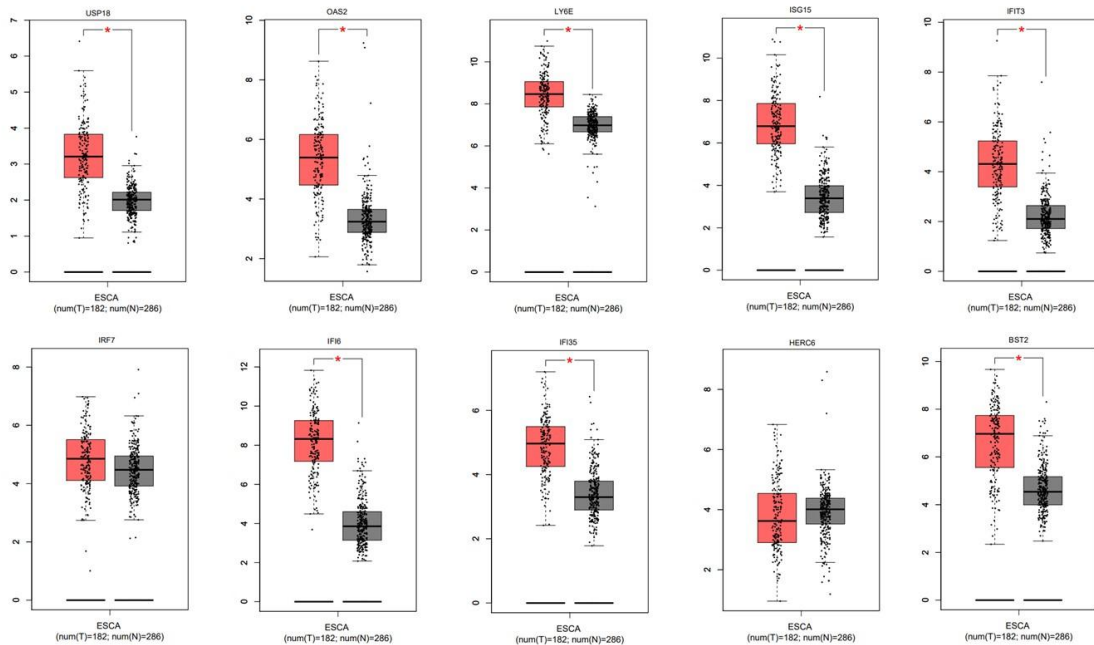


211
212 **Figure. 3:GO and KEGG enrichment analysis for common DEGs across all stages of oral**
213 **cancer.** The figure shows enriched biological processes and pathways, with bar graphs
214 depicting the significance of each term, highlighting key functional categories and pathways
215 associated with these DEGs.

216 Validation of identified potential genes in Esophageal carcinoma

217 To validate the potential clinical significance of the identified genes, we further investigated
218 their differential expression in Esophageal carcinoma (ESCA) using the GEPIA platform
219 (<http://gepia.cancer-pku.cn/>). Leveraging the robust datasets of TCGA and GTEx, comprising
220 182 tumors and 286 normal liver tissue samples, we implemented stringent criteria of absolute
221 \log_2 fold change ($|\log_2FC| \geq 1$) and p-value < 0.01 to identify differentially expressed genes
222 (DEGs) with high confidence.

223 This rigorous approach yielded a comprehensive landscape of gene expression alterations
224 associated with OSCC progression. Tumor samples, visually distinguished by the red color on
225 the GEPIA platform, exhibited distinct expression patterns compared to their grey-hued normal
226 counterparts. Significant biomarkers are marked with a red star sign in the box plot of
227 expression analysis, which is for the genes USP18, OAS2, LY6E, ISG15, IFIT3, IFI6, IFI35,
228 and BST2. At the same time, insignificant markers are IRF7 and HERC6 (Fig. 4). This stark
229 contrast highlights the altered transcriptional activity within the cancerous context, providing a
230 valuable starting point for further investigation.

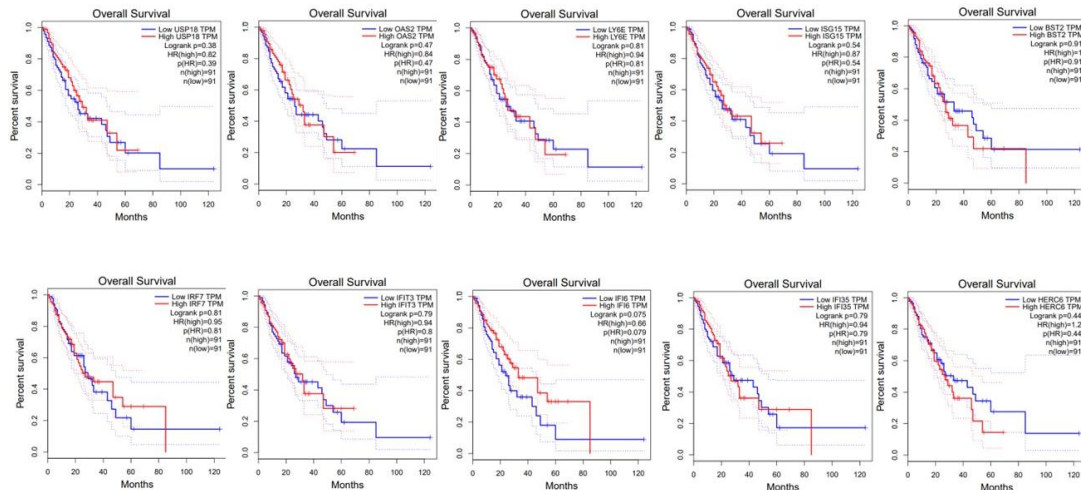


231
 232 **Figure 4: Box plot showing the distribution of expression levels for significant genes across**
 233 **different conditions or groups. Each box represents the range and median of gene expression,**
 234 **highlighting differences in expression between groups and identifying outliers.**

235 Potential Gene-driven Survival Analysis

236 In the comprehensive survival analysis, the hazard ratios for specific genes signify their roles
 237 in the progression of diseases, with the summarized findings presented in Figure 5. Elevated
 238 expressions of USP18 (HR= 0.82) and OAS2 (HR= 0.84) correlate with increased mortality
 239 risks, underscoring their significance in cancer development, where USP18 affects interferon
 240 signaling and immune responses against tumors, while OAS2, known for its antiviral
 241 properties, also plays a role in inhibiting tumor growth. ISG15 (HR= 0.87) and IRF7 (HR=
 242 0.95) are linked to higher mortality rates, highlighting their contributions to cancer cell
 243 proliferation; ISG15's dual role in tumorigenesis and IRF7's involvement in immune response
 244 modulation further elaborate their complex interactions in cancer dynamics. IFI6 emerges with
 245 a hazard ratio of 0.66, hinting at a protective role, possibly through its anti-apoptotic functions,
 246 which might oppose tumor development in certain contexts. The genes IFIT3 (HR= 0.94),
 247 IFI35 (HR= 0.94), and BST2 (HR= 1) show hazard ratios close to 1, indicating a more neutral
 248 prognostic value in this study. Yet, their biological functions—ranging from immune signaling
 249 to protein degradation pathways—offer avenues for further cancer-related inquiries. HERC6,
 250 with an HR of 1.2, suggests a slightly increased risk of mortality linked to its role in protein
 251 degradation pathways, which, when altered, can contribute to cancer. LY6E's expression (HR=
 252 0.94) suggests a potential risk in cancer progression, possibly through its impact on cellular

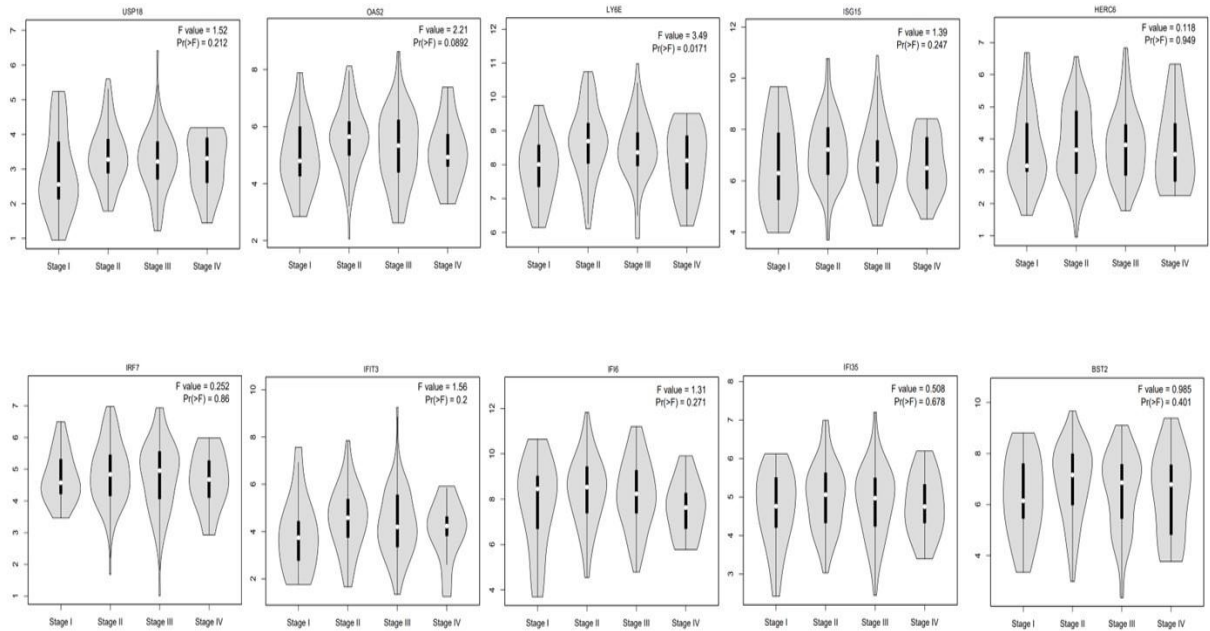
253 immunity and response mechanisms. This indicates a need for further exploration in oral
254 squamous cell carcinoma (OSCC) and other cancers Fig. 5, Table 1.



255
256 **Figure 5: Survival plot illustrating the association between the expression levels of**
257 **significant genes and patient survival.** The plot shows survival curves for different expression
258 groups, highlighting how high or low expression of these genes correlates with survival
259 outcomes.

261 Gene expression analysis across cancer stages revealed distinct patterns

262 The violin plots are based on the Esophageal carcinoma (ESCA) data using the GEPIA
263 platform (<http://gepia.cancer-pku.cn/>). Leveraging the robust datasets of TCGA and GTEx,
264 comprising 182 tumors and 286 normal liver tissue samples, we implemented stringent criteria
265 of absolute log2 fold change ($|\log_2FC| \geq 1$) and p-value < 0.01 to identify differentially
266 expressed genes (DEGs) with high confidence. It illustrates the distribution of gene expression
267 levels across different cancer stages for ten genes implicated in OSCC. While some genes
268 exhibited consistent trends, others demonstrated more complex patterns. USP18 and OAS2
269 displayed a progressive upregulation with advancing disease stages, suggesting their potential
270 roles as oncogenes. Conversely, IFI6 showed a distinct pattern with higher expression in early-
271 stage disease, hinting at a potential tumor-suppressive function. ISG15 and IRF7 exhibited
272 more variable expression profiles, necessitating further investigation to elucidate their precise
273 roles. The remaining genes, LY6E, IFIT3, IFI35, BST2, and HERC6, showed less pronounced
274 expression changes across stages, suggesting their potentially limited impact on disease
275 progression (Fig. 6, Table 1).



276
 277 **Figure 6: Violin plot displaying the distribution of expression levels for significant genes**
 278 **across different stages.** Each violin shape represents the density and distribution of gene
 279 expression within each stage, allowing visualization of variation and central tendencies.

280
 281 **Network analysis reveals interconnectedness of key biomarkers in gallbladder cancer**
 282 Network analysis reveals the interconnectedness of key biomarkers in OSCC. Building upon
 283 the identification of potent prognostic LY6E gene in OSCC, we delved deeper into their
 284 regulatory networks using NetworkAnalyst, a powerful tool for exploring biological
 285 interactions depicted in Fig. 6. This comprehensive analysis revealed a complex interplay
 286 between transcription factors (TFs), microRNAs (miRNAs), and these key genes, shedding
 287 light on the molecular mechanisms underlying their impact on patient survival. LY6E emerged
 288 as a central hub, with interactions identified for 7 TFs, 26 miRNAs, and 1 additional gene. This
 289 extensive network suggests a multifaceted regulatory landscape where LY6E's influence on
 290 survival likely involves diverse transcriptional and post-transcriptional mechanisms.

313 study, which employs a network-centric approach, unveils the complex interactions among
314 differentially expressed genes in OSCC, highlighting the crucial role of LY6E in driving the
315 disease's development and prognosis. Further understanding of this molecular interplay could
316 lead to the design of innovative therapeutic strategies to improve patient outcomes in the fight
317 against OSCC. Nevertheless, conducting studies on a larger patient cohort and in-depth basic
318 mechanistic studies would provide more detailed information and implications of the LY6E
319 gene. Our study sets the stage for future detailed investigations on LY6E as a potential
320 biomarker for the early diagnosis of OSCC.

321 **DECLARATION:**

322 **Acknowledgment**

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324 **Author's contributions**

325 SBP: conceptualization and prepared the outline of the research article. RKG, RB, and HI
326 wrote the manuscript: RB, and RKG prepared figures and tables for the manuscript. DK has
327 done referencing and data analysis. All authors read, reviewed and approved the submitted
328 version.

329 **Availability of data and materials**

330 Not applicable

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332 None

333 **Conflicts of interest**

334 All authors have showed no conflicts of interest.

335 **Ethical Statements:** This article does not contain any studies with human participants or
336 animals performed by any of the authors.

337 Authors also declare that 'Clinical trial number: not applicable.'

338 **Consent for publication**

339 Not applicable.

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