

# 1 Optimizing Sanitization and Access Control: Lessons from a Cleaning Validation Study.

## 5 1. Abstract

6 **Background:** Cleaning Validation studies is an important aspect in pharmaceutical studies to identify and correct  
7 potential problems previously unsuspected, which could compromise the safety, or quality of subsequent batches of  
8 product produced within the equipment..  
9

10 **Methods** The study was designed as experimental study and conducted between January-June 2025.The study was  
11 conducted on Clobetasol 0.05% cream manufacturing equipments.Swabbing technique was used to find microbial  
12 load on equipment under the study. Total aerobic Microbial Count (TAMC) 100 cfu/ 100 cm<sup>2</sup> and Total Yeast and  
13 Mould Count 10cfu/ 100 cm<sup>2</sup> was set as pass limit during cleaning validation study. Different contamination  
14 variables were studied during cleaning validation study.

15  
16 **Result:** The mean value of cleaning validation study for Total aerobic Microbial count and Total yeast and mould  
17 count was found to be 8.33 and 0.55 for all three batches of used equipments respectively. Standard deviation was  
18 found to be 6.08 and 0.511, obtained result was not found statistically significant with respect to p- value <0.05.

19 **Conclusion:** Several contamination variables like sanitization technique, frequency of fumigation, access control in  
20 area, Microbial load over lint free clot used for mopping was coined during study and after mitigating such  
21 contamination variables the study result was found satisfactory.Changed technique like change from chemical  
22 method of sterilization to moist heat sterilization of templates , use of lint-free cloths soaked in 70% IPA provided  
23 better control, Increased fumigation frequency and limited access in area acts as major preventive measures for  
24 contamination Control.

25 Key Words: Cleaning Validation, Contamination Variables, Findings, Swab technique, Root cause analysis  
26

## 27 2.Introduction

28 Cleaning validation is study done to find the effectiveness and reliability of cleaning pharmaceutical production  
29 equipment. People in the pharmaceutical industry use equipment validation and cleaning procedures mainly to  
30 prevent cross-contamination that makes these practices crucial.[1]In general, cross-contamination usually happens  
31 when an active ingredient from one product is transferred to other product through the instruments improper  
32 cleaning that can pose real risks to consumers and second type is contamination by foreign materials, which could be  
33 bacteria or fungi .Poor maintenance or storage conditions may let microbes flourish in processing equipment and  
34 becomes a serious issue[2]

35 The most important benefit of conducting cleaning validation work is to identify and correct potential problems  
36 previously unsuspected, which could compromise the safety, efficacy or quality of subsequent batches of drug  
37 product produced within the equipment.[3]

38 Cleaning validation guarantees the safety, identity, purity, and strength of products, which are fundamental to cGMP  
39 (Current Good Manufacturing Practice) [4]

40 Significance of Cleaning Validation study

41 The primary purpose of cleaning validation is to prevent cross-contamination between different pharmaceutical  
42 products, thereby ensuring product integrity and patient safety. By verifying the effectiveness of cleaning  
43 procedures, it provides assurance that active ingredient or cleaning agent from a previous batch remains under  
44 acceptable level on equipment that could not adulterate subsequent products[5].The process typically includes the

45 development of cleaning methods, assessment of worst-case scenarios, establishment of acceptance criteria, and  
46 verification using appropriate analytical techniques to quantify residuals and ensure reproducibility [6].

47 Cleaning Method Development [8,9]

48 Cleaning method development is a critical component of the pharmaceutical validation lifecycle and is carried out  
49 alongside drug development to ensure that cleaning processes are scientifically sound, efficient, and compliant with  
50 regulatory expectations. Stages of Cleaning Method Development Cleaning method development typically  
51 progresses through three main stages:

52 1. Feasibility: This stage evaluates whether the proposed method is suitable for the specific sample, equipment, and  
53 contaminants in question.

54 2. Development: Optimization of cleaning parameters such as time, temperature, cleaning agents, and equipment to  
55 achieve maximum residue removal.

56 3. Validation: Demonstration that the optimized cleaning method consistently meets acceptance criteria across  
57 multiple runs and conditions

58 Risk-Based Approach

59 A risk-based strategy is fundamental in cleaning validation studies. It involves identifying and prioritizing  
60 equipment, surfaces, or products that pose a higher risk of contamination or cross-contamination. This approach  
61 allows the validation team to focus resources and attention on critical areas where product safety or quality may be  
62 most vulnerable. It involves in choosing appropriate cleaning method, test method and justify the result based on  
63 regulatory guidelines. Risk-based approaches include Failure Mode and Effects Analysis (FMEA), Fault Tree  
64 Analysis (FTA), Hazard Analysis and Critical Control Points (HACCP), and Quantitative Microbiological Risk  
65 Assessment (QMRA) [10]

66 Scope of study

67 • To study different variables for contamination of clean equipments

68 • To provide scientific rationale and documentation for cleaning effectiveness.

69 • To perform cleaning validation study on equipments used for formulation of products

### 70 3.Methods

#### 71 3.0:Study Design

72 The study was designed as experimental study and conducted between January-June 2025.The study was conducted  
73 on Clobetasol 0.05% cream manufacturing equipments in class D manufacturing facility in pharmaceutical  
74 Industry.Sampling method was selected as Swabbing technique, to study the microbial load on equipments included  
75 the study.Total aerobic Microbial Count (TAMC) 100 cfu/ 100 cm<sup>2</sup> and Total Yeast and Mould Count 10cfu/ 100  
76 cm<sup>2</sup> was set as pass limit during cleaning Validation study. [12].

#### 77 Study was conducted in 3 phases

78 a.To study the actual status of microbial load on clean equipments under study

79 b.To find the different variables for contamination of clean equipments

80 c.To perform cleaning validation study on equipments used for formulation of products

81 Test conditions of cleaning validation was designed under static condition.Temperature less ( $\leq 25$  °C), and Humidity  
82 ( $\leq 60\%$ ) was maintained in sampling areas during study periods.Swabbing technique was used for sampling and  
83 membrane filter test method was used for detection of microbial load on equipments included in the study.

84 **3.2: Materials Used under study**

85 Materials and Manufacturer

86 1 Buffered peptone water: Hi Media

87 2 Soyabean Casein Digest Agar (SCDA): Hi Media

88 3 Sabouraud Dextrose Agar (SDA): Hi Media

89 4 Sterile swab: Hi Media

90 5 Autoclave: Equitron

91 6 Bio-safety Cabinet:Thermolab

92 7 Incubators:Allyone

93 8 Colony Counter: Lapid

94 9.70% IPA: Qualigens

95

96 **3.3: Sampling Procedure**

97 Test areas of  $10 \times 10 \text{ cm}^2$  were measured using sterile stencils. The sterile swabs were moistened with sterile water,  
98 and samples were collected from two different  $100\text{cm}^2$  areas of each piece of clean equipment. A total of two swab  
99 samples from each equipment were collected using unidirectional movements—first 10 horizontal Strokes followed  
100 by 10 vertical strokes—for the determination of total aerobic microbial count and total yeast and mould count. The  
101 swabs were placed into separate test tubes containing 10 mL of Buffered peptone water (Fig.1) and transported to  
102 the microbiology laboratory.



Fig 1: Swab Sample in Buffered peptone water

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105 **3.4 Sample analysis and Quality Control**

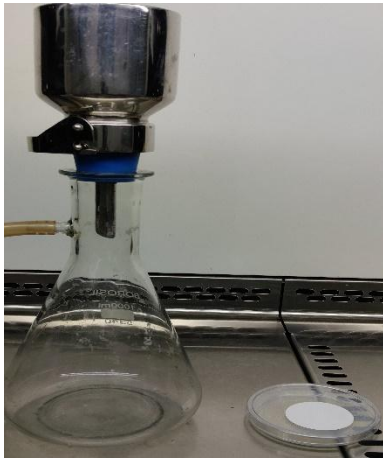
106 The equipment used during the study was well calibrated. Stencils used for measuring surface area were sterilized in  
107 an autoclave at  $121 \text{ }^\circ\text{C}$  and 15 psi for 15 minutes. Soybean Casein Digest Agar was used for the isolation of bacteria,  
108 and Sabouraud dextrose agar with chloramphenicol was used for the isolation of fungi. Growth promotion test and  
109 Sterility checks of the swab sticks was performed as per USP<61> (United States Pharmacopoeial Convention)  
110 2025[11]. Each tube containing the swab sample was shaken for 2-3 minutes. 10 ml of the sample solution was  
111 individually pipetted into 50 mL of peptone water, mixed thoroughly, and the entire contents were filtered through a  
112 membrane filter with a pore size of  $0.45 \text{ }\mu\text{m}$ . (Fig.2)



113

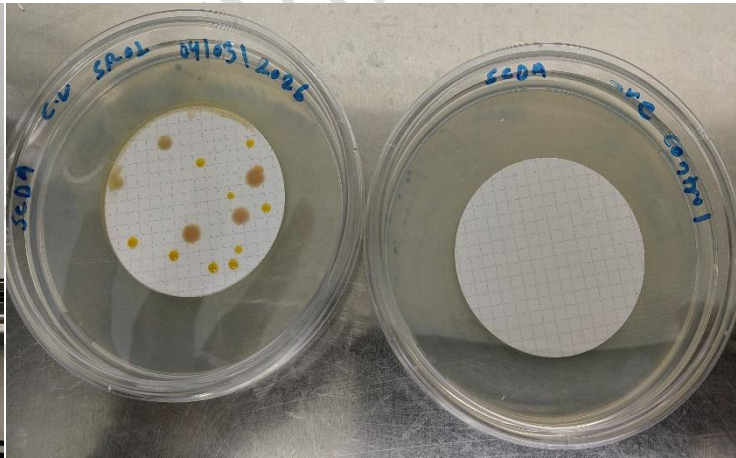
114 Fig.2: Filter Assembly,Swab sample and Buffered Peptone water,

115 The membrane filter was aseptically transferred onto Soybean Casein Digest Agar (SCDA) using sterile forceps  
116 (Fig.3) and incubated at 35 °C for 72 hours. For total yeast and mould count, the filter was placed on Sabouraud  
117 Dextrose Agar with chloramphenicol plates and incubated at 25 °C for 5 days [7].The microbial growth was counted  
118 using colony counter and reselt were interpreted.(Fig.4)



119

120 Fig.3: Filtered sample on agar plate



Fi.g 4: Microbial growth on Sample after Incubation

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#### 4.0 Result

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##### 4.1: Initial Result of cleaning validation study

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The load of Microorganism (Total aerobic microbial count) was found out of limit (ie.> 100 Cfu/100cm<sup>2</sup>) and the total yeast and mould count was found with in pass limit;<10 Cfu/100cm<sup>2</sup>.. During the disinfection process of equipments, final rinse was done with purified water. Then 70% IPA was sprayed and mopped with cotton cloth. The result showed that multiple factor may have contributed to the increase bacterial load of on clean equipments under study (Table 1)

**Table1: Initial Microbial Load on equipments**

S.No.	Equipment	TAMC(Cfu/100cm <sup>2</sup> ) (Limit:<100 cfu)		TYMC(Cfu/100cm <sup>2</sup> ) (Limit:<10 cfu)	
		Sampling Location	Result	Sampling Location	Result
1	Wax vessel	From the both side of the baffles (MC-01)	300	From the base of the vessel near discharge(Mc-02)	1
2	Manufacturing Vessel	A sample from the Teflon flanges attached to the homogenizer. (MC-03)	250	A sample from the base of the equipment near the drain pipe(MC-04)	3
3	Storage Vessel/Paste preparation vessel	Sample from the Base on the other (MC-05)	150	Sample from the wall of the vessel (MC-06)	1
4	SS Containers	Wall of the container(MC-07)	180	Base of the container(MC-08)	2
5	SS Jugs	Base of the jugs(MC-09)	200	Base of the jugs(MC-10)	3
6	Semi- Automatic Tube Filling Machine	Inner surface of hopper(MC-11)	240	Surface of the stirrer(MC-12)	4

131 **4.2:Root cause Analysis to find source of contamination**

132 Different variables was selected to find the source of increase microbial load on clean.Test of the total yeast and  
133 mould count was excluded in the study as the result was found satisfactory (Table 1)

134 1. Sample taken without any disinfection process

135 2.Spray with 70% IPA directly on clean equipment, Air Dry

136 3. Spray 70% IPA on Lint free cloth and mop the clean equipment

137 4. Completely dip Lint free Cloth in 70% IPA and Mop the clean equipment

138 5. Mop the template with 70% IPA and swab taken

139 6. Lint free cloth soaked in sterile water and mopped the clean equipment

140 7.Limiting personnel access in the sampling area

141 8.Decreasing frequency of fumigation in the area

142 **Table 2: Observation of different variables of Study for Microbial load**

S.No.	Variables	TAMC (Cfu/100cm <sup>2</sup> ) (Limit:<100 cfu)
1	Sample taken without any disinfection process	190
2	Spray with 70% IPA directly on clean equipment, Air Dry	46
3	Spray 70% IPA on Lint free cloth and mop the clean equipment	330
4	Completely dip Lint free Cloth in 70% IPA and Mop the clean equipment	21
5	Mop the template with 70% IPA and swab taken	10
6	Lint free cloth soaked in sterile water and mopped the clean equipment	1300

143

144 The result showed that lint free cloth as main carrier of organism to the clean equipments during clean validation  
 145 study and also the chemical sanitizing technique of template was not effective.(Table 2)

146 **4.3:Remediation of Microbial Contamination on Clean Equipments**

147 **Changed Technique**

148 1. Template sterilization technique was changed from chemical sterilization to moist heat sterilization (Autoclaving)

149 2. Cloth used for moping was dipped in 70% IPA and Dried before use of final moping on clean equipments.

150 3. Fumigation frequency in the area was decreased to 3 month to 1 month.

151 4. Limited access was done in area during sampling in the area.

152

153 **Table 3:Result after correction of the contamination variables**

S.No.	Equipment	Sampling Location	TAMC (Cfu/100cm <sup>2</sup> ) (Limit:<100 cfu)
1	Wax vessel	From the both side of the baffles (Mc-01)	7
2	Manufacturing Vessel	A sample from the Teflon flanges attached to the homogenizer (MC-03)	9
3	Storage Vessel/Paste preparation vessel	Sample from the Base on the other (MC-05)	23
4	SS Containers	Wall of the container(MC-07)	30
5	SS Jugs	Base of the jugs(MC-09)	20
6	Semi- Automatic Tube Filling Machine	Inner surface of hopper(MC-11)	17

154

155 After correction of the contamination Variables, the result of Total Aerobic microbial count was found in pass limit  
 156 < 100 Cfu/100cm<sup>2</sup>.(Table 3)

157 **Table 4:Cleaning Validation Study Result**

S.No.	Equipment	TAMC(cfu/100cm <sup>2</sup> ) (Limit:<100 cfu)				TYMC(cfu/100cm <sup>2</sup> ) (Limit:<10cfu)			
		Sampling Location	Result			Sampling Location	Result		
			Batch1	Batch2	Batch3		Batch1	Batch2	Batch3
1	Wax vessel	From the both side of the baffles(Mc-01)	7	1	9	From the base of the vessel near discharge (Mc-02)	0	1	0
2	Manufacturing Vessel	A sample from the Teflon flanges attached to the homogenizer (MC-03)	10	3	2	A sample from the base of the equipment near the drain pipe(MC-04)	1	0	1
3	Storage Vessel/Paste	Sample from the Base on the	6	3	2	Sample from the wall of the	0	1	1

	preparation vessel	other(MC-05)				vessel (MC-06)			
4	SS Containers	Wall of the container(MC-07)	20	23	14	Base of the container (MC-08)	1	0	1
5	SS Jugs	Base of the jugs(MC-09)	10	12	8	Base of the jugs(MC-10)	0	1	0
6	Semi-Automatic Tube Filling Machine	Inner surface of hopper(MC-11)	5	11	5	Surface of the stirrer(MC-12)	1	0	1
Mean value			8.38			Mean value	0.55		
Standard Deviation			6.08			Standard Deviation	0.511		

158

159 The result of cleaning validation on Clobetasol 0.05% cream manufacturing equipments in class D manufacturing  
160 facility in pharmaceutical Industry shows within the acceptance limit below 100 cfu and 10 cfu respectively for Total  
161 aerobic microbial count and Total Yeast and Mould count, after correcting the contamination variable.(Table 4)The  
162 mean value of Total aerobic Microbial count and Total yeast and mould count was found to be 8.33 and 0.55 for all  
163 three batches of used equipments respectively.Standard deviation was found to be 6.08 and 0.511, obtained result  
164 was not found statistically significant with respect to p- value <0.05.

165 Discussion

166 Cleaning Validation studies is an important aspect in pharmaceutical studies to identify and correct potential  
167 problems previously unsuspected, which could compromise the safety, efficacy or quality of subsequent batches of  
168 drug product produced within the equipment. During the study we studied the potential contaminating variables and  
169 mitigate the causes to guarantees the safety, identity, purity, and strength of products, which are fundamental to  
170 cGMP during formulation of products. Several contamination variables like inappropriate disinfection techniques,  
171 use of lint free cloth with higher microbial load, fumigation time interval, unauthorized personnel assess in the area  
172 were found as major variable which increased microbial load on clean equipments.(Table2)

173 After change in techniques like, Template sterilization technique was changed from chemical sterilization to moist  
174 heat sterilization (Autoclaving), Cloth used for moping was dipped in 70% IPA and Dried before use of final  
175 moping on clean equipments, Fumigation frequency in the area was decreased to 3 month to 1 month and Limited  
176 access was done in area during sampling in the area the result of cleaning validation was found satisfactory (Table 3)

177 Cleaning validation study on Clobetasol 0.05% cream manufacturing equipments was performed after correcting the  
178 different contamination variables.The mean value of obtained microbial count was found to be 8.33 and 0.55 for all  
179 three batches of used equipments respectively (Table 4) but the obtained data was was not found statistically  
180 significant with respect to p- value <0.05.The lower microbial load on equipments might be due to correction in  
181 contamination variables found during study (Table 3).Major Root cause of contamination found during study,  
182 include technique of sterilization, personnel access in area, duration of Fumigation in area and the carryover of  
183 microbial load from lint free cloth to clean equipments during sanitization of equipments.

184 Conclusion

185 1. The high Total aerobic microbial count was likely caused by poor sterilization of the sampling tools  
186 (Template/Cloth) rather than the equipment cleanliness itself.

187 2. The change from chemical method of sterilization to moist heat sterilization of templates was critical.

188 3. The use of lint-free cloths soaked in 70% IPA provided better control.

189 4. Increased fumigation frequency and limited access improved the environmental control

190 **Conflicts of Interest**

191 The authors declare that they have no competing interests that could have influenced the objectivity or outcome of  
192 this research.

193  
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200  
201 **Informed consent**

202 This study did not involve human participants, human data, or human tissue. Therefore, approval from an IRB or  
203 ethics committee was not required. All procedures were conducted in accordance with the relevant guidelines and  
204 regulations for laboratory-based research.

205  
206 **Large Language Model**

207 No large language model was used in the preparation of this article

208  
209 **Authors Contribution**

210 SKS contributed to conceptualization, methodology, investigation, data collection, analysis, and writing of the  
211 original draft and final manuscript.

212  
213 **Data Availability**

214 No datasets were generated or analyzed for this study beyond the summary results presented in the article; therefore,  
215 no additional data are available. Data sharing does not apply to this research. For reasonable queries about the  
216 summarized results or methods, please contact the corresponding author listed in the manuscript.

217  
218 **Ethics Approval and Consent**

219 No human or animals samples were involved in the study,so no ethical approval and consent is required in this  
220 study.

221  
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