

# 1 **Bio-control of Raw Diatomaceous Earth alone or/in combination with permethrin dust** 2 **against *Rhyzopertha dominica*(Coleoptera: Bostrychidae) on stored wheat (*Triticum*spp)**

## 3 **Abstract**

4 Laboratory experiments was carried out to evaluate the efficacy of raw DE alone or/in combination with  
5 permethrin dust against *R. dominica* on wheat, to assess insect mortality caused by exposure to raw DE and/ or  
6 permethrin, effects on progeny production of exposed beetles and prevention of grain damage by the test insect.  
7 *R. dominica* was obtained from laboratory cultures which has been maintained in the laboratory for over year,  
8 where F<sub>1</sub> progeny was used for the experiment. The raw DE was tested at application rates of 0 ( untreated  
9 control) 500, 750, 1000, 1500 mg raw DE/kg alone and with 2 and 5 mg active ingredient permethrin to each  
10 DE dose making a total of 12 treatment combinations. Each treatment combination and control, 50 g grain  
11 samples in three replicates were place in 250 ml capacity bottles, and thirty adults insects were placed into each  
12 replicate. Adult mortality, progeny production and percentage damage kernel by *R. dominica* were assessed. The  
13 result reveals that at higher dose rate of 1500 mg/kg of raw DE and with combination of permethrin gave  
14 appreciable adult mortality after 7 days of exposure interval, after continuous exposure to 14 days, complete  
15 100% adult mortality was noticed at 1500 mg/kg dose rate. Progeny production was suppressed after 40 and 80  
16 storage period. Greater than88% progeny suppression where recorded at the lowest dose rate of 500 mg/kg on  
17 raw DE and enhanced DE as the dose rate increases, complete progeny inhibition were noticed at the highest  
18 dose rate of 1500 mg/kg or 500 mg + 2 or 5 mg permethrin when compared with untreated controls. The  
19 percentage damage kernel decreases at the dose rate of 1500 mg/kg with raw DE combined with 2 and 5 mg/kg  
20 were 1.3±1.0 and 0.0±0.0, respectively.

21 **Key words:** Wheat Diatomaceous earth *Rhyzoperthadominica* Mortality Progeny

## 22 **Introduction**

23 Wheat *Triticum spp* is one of the most important cereal in the world. It is staple food  
24 for humans and livestock, it is of prime importance when compared to other cereals (Salman  
25 and Hamad, 2024). Wheattrading occuppies major share in world market when compared to all  
26 other cereal crops combined it supersedes rice and maize in its source of protein and calories  
27 (Mostafa and Abotaleb, 2024). Wheat production, is hindered by numerous factors which  
28 cause quantitative and qualitative losses both before and after harvest (Susurluk and Bütüner,  
29 2024). One of the primary factors contributing to these losses is pests (Mahroofet *al.*, 2010).  
30 Lesser grain borer, *Rhyzopertha dominica* (F.), is one of the destructive primary insect pests  
31 which infests wheat and many cereal grains at post-harvest levels (Naseem and Khan, 2011;  
32 Edde, 2012). It is usually a polyphagous and cosmopolitan insect pest all over the world  
33 (Majeed *et al.*, 2015; Suleiman and Rugumamu, 2017). *R. dominica* infestations have been  
34 reported to have reduced grains to dust (Salmanet *al.*, 2024).

35 *R. dominica*larvae consume both germ and endosperm during their development in  
36 grain and provide more frass and the feeding on seed germ reduces germination rates and  
37 vigour of the grains and may be followed by secondary pests and fungi (Manivannanet *al.*,  
38 2024). This insect is capable of damaging grain, causing weight losses of up to 40%  
39 (Baliotaet *al.*, 2024). Such damage can be quite serious economically (Cao *et al.*, 2024). It is  
40 reported that *R. dominica* infestation on wheat grains resulted in substantial change in the  
41 nutritive contents such as calcium, phosphorus, zinc, iron, copper, manganese and total  
42 content of lipids, phospholipids, galactolipids, polar and non polar lipids and vitamin content  
43 of the wheat grains (Jood *et al.*, 1996; Perišić *et al.*, 2018; Milosavljevicet *al.*, 2024), and  
44 reduction in the starch digestibility (Dhaliwal *et al.*, 2010; Kerbelet *al.*, 2024). Infestation of  
45 *R. dominica*on wheat grains cause losses of 23 to 29% thiamine, 13 to 18% riboflavin and 4  
46 to 14% niacin (Iqbal *et al.*, 2024).

47 *R. dominica* is indeed a notable pest in the realm of grain storage. This species  
48 presents a major risk to stored grains like corn, wheat, rice, and sorghum in numerous  
49 regions globally (Erturket *et al.*, 2024). Both adult and larvae of *R. dominica* contribute to the  
50 deterioration of stored grain quality as they feed (Arthur *et al.*, 2012; Edde, 2012). The life  
51 cycle of *R. dominica* begins with adult females laying eggs shortly after mating (Hill, 2003;  
52 Mortazaviet *et al.*, 2024), each female can lay 300 to 500 eggs during her reproductive period  
53 (Deshwal *et al.*, 2018). Once the eggs are hatch, the larvae begin their destructive feeding  
54 habits. They penetrate grains or consume flour and small particles that escape from holes  
55 created by adults (Cilgin and Kececi, 2024). Adult *R. dominica* also exacerbates the damage  
56 by entering grains to feed and continue developing (Mahmood *et al.*, 2024). This continuous  
57 cycle of egg laying, larval feeding, and adult feeding allows *R. dominica* to potentially  
58 complete 3 to 4 generations per year under favorable conditions (Majeed *et al.*, 2015;  
59 Javanmard *et al.*, 2023).

60 Synthetic chemical insecticides have been employed for many years as a control  
61 method to manage pests, and their use persists today (Susurluk and Bütüner, 2024). However,  
62 recent European Union (EU) decisions have led to barred on use of chemical insecticides in  
63 agricultural production, particularly in stored productspest management (Deshwalet *et al.*,  
64 2018). Some of the main reasons for these restrictions are the harmful effects insecticides on  
65 non-target organisms and the residues they leave on stored products (Wakilet *et al.*, 2024). This  
66 decision has increased the quest for alternative insect pest management methods over  
67 chemical control. However, findings have shown that the use of Diatomaceous earth which  
68 have little or no side effects as compared to the problems pose to the stored products using  
69 synthetic chemical insecticides. DEs acts as physical protectants against insect pests by  
70 inhibiting reproduction, growth or development through exoskeleton abrasion, cuticle  
71 piercing and absorption of fluid thereby dehydrating the insect pest (Majeed *et al.*, 2015).  
72 Raw DEalone/or in combination with other formulations confer effective insect pest control  
73 in stored grains. DEs have been used as physical insecticides for thousands of years by  
74 earliest peoples and are presently used in modern grain storage facilities(Susurluk and  
75 Bütüner, 2024). The objectives of this study was evaluate insecticidalpotential of raw  
76 diatomaceous earth alone or in combination with permethrin dust in the management of *R.*  
77 *dominica* on stored wheat.

78

## 79 **Material and Methods**

80 The Experiment was conducted at the Entomology Laboratory of the Department of  
81 Crop Protection, Faculty of Agriculture, University of Maiduguri, Nigeria. Latitude 11<sup>0</sup> 50'  
82 42" N, 13<sup>0</sup> 9' 36" E. Temperature and relative humidity during the study was measured using  
83 hygrometer. Temperature and relative humidity during the study were range from 34<sup>0c</sup> and  
84 65-75 % relative humidity.

### 85 **3.1 Sources of Experimental Materials**

86 The insects *R. dominica* was obtained from laboratory cultures which has been  
87 maintained for over a year. The wheat variety *cetia* was obtained from Lake Chad Research  
88 Institute, Maiduguri. The DE was supplied in a form of crude soft chalky rock. The rock was  
89 milled fine in the laboratory using a pestle and mortar and pass through a fine sieve (150 $\mu$ ) to  
90 obtain a powdery consistency. The fine powder was analyzed for  $p^H$  and tapped density in  
91 accordance with method describe by Korunic (1997). While mineral composition were  
92 analyzed in the Mineralogy Laboratory of the Department of Geology, University of

93 Maiduguri by X-ray florescence method on Minimate (Panalytical Company, UK). The  
94 insecticide permethrin dust (0.6% a.i). Manufactured by Gongoni Company Limited Kano,  
95 Nigeria was obtained and used for the experiment.

### 96 **3.2 Preparation of Wheat Grains**

97 The wheat grains used for the experiment were cleaned, disinfested and placed in 2 kg  
98 plastic containerwith cover was used until commencement of experiments.

### 99 **3.3 Insects Culturing Procedures**

100 *R. dominica* was cultured on wheat. For this purpose 200 unsexed insects were placed  
101 on 500g wheat in 1 liter capacity jar and then the parents were removed after 15 days of  
102 feeding and oviposition. Parent insects were removed by emptying the content of the wheat  
103 on to plastic tray and removing the adult insects from the wheat and returning the grains and  
104 grain dust to the culture jar. The resulting F<sub>1</sub> progeny were used for test. The stocks were  
105 maintained under ambient laboratory conditions.

### 106 **3.4 Grain Treatment.**

107 The grain was treated at dose rate of 0 (untreated control) 500, 750, 1000, 1500mg  
108 raw DE/Kg, alone and with 2 and 5mg active ingredient (a.i) permethrin to each DE dose  
109 making a total of 12 treatment combinations. For each treatment combination 150g of wheat  
110 grain was placed in 1 liter capacity bottles and different levels of DE and /or permethrin dust  
111 were added. The bottles were closedwith their lids and shaken manually for 300 seconds (5  
112 minutes) to acquire uniform distribution on the entire wheat grain. Same quantities of grain  
113 were kept untreated to serve as control.

114

### 115 **3.5 Bioassay Procedures.**

116 Each treatment combinations and control, 50g wheat grain samples in three replicates  
117 was placed in 250ml capacity bottles, and thirty (30) adult insects were placed in each  
118 treatments replicate. Adult mortalities were observed after 7 and 14 days exposure and each  
119 bottle was emptied on to a tray, live and dead insects were recorded and dead insect were  
120 removed from the bottles, and the live insects were returned into the bottles and recounted  
121 after 14 days of exposure, at this time both the live and dead insects were removed from each  
122 treatment bottles.

123 The grain and the grain dust were returned to their appropriate bottles and kept on the  
124 laboratory shelf at the same conditions, for additional 40 and 80 days intervals for adult  
125 progenies emergence of F<sub>1</sub> and F<sub>2</sub>, respectively.

### 126 **3.6 Experimental Design and Data Analysis**

127 The experiment was laid in split-split-plot design. Insect specie was assigned to the  
128 main plot, while exposure period and dose rate were assigned to the sub-plot and sub-subplot,  
129 respectively. Data on adult mortality was expressed in percentage and the data were corrected  
130 for mortality in control using the Abbott's formula (Abbott, 1925):

$$131 P_T = (P_o - P_c / 100 - P_c) 100$$

132 Where, P<sub>T</sub> - Corrected mortality;

133 P<sub>o</sub> - Observed mortality;

134 P<sub>c</sub> - Mortality in control

135 Data on progeny and grain damage were log (log<sub>x+1</sub>) transformed and subjected to  
136 ANOVA with number of progeny and percent grain damage as response to variables and dose  
137 rate as main effects. For comparison of treatment means, the Tukey-Kramers' HSD test at  
138 P≤0.05, was used.

139

## 140 **4.0 Results**

141 **4.1 Effect of raw and enhanced DE on mortality of *R. dominica* and adults after 7 days**  
 142 **of exposure.**

143 The results obtained shown that DE and permethrin alone or in combination can cause  
 144 high mortality in *R. dominica*. Significant differences ( $p < 0.05$ ) in adult mortality were  
 145 observed among different dose rates of both DE alone and in combination with permethrin.  
 146 Increased with increase in dose rate in all cases causes adult mortality of the test insect. Raw  
 147 DE at a rate of 500 mg /kg gave  $65.7 \pm 4.7$  mortality of *R. dominica* after 7 days of exposure,  
 148 while as the dose rate increases, the rate of mortality also increases. At 1500mg/ kg mortality  
 149 was  $81.1 \pm 3.1$  of *R. dominica*, also at 7 days of exposure (Table 1). As raw DE combined with  
 150 2 mg/kg permethrin result in increased beetle mortality. A combination of 2 mg/kg  
 151 permethrin and 500 mg/kg of raw DE after seven days of exposure resulted in mortality rates  
 152 of  $66.7 \pm 3.9$  of *R. dominica*. However, as the DE dose rate increases, the mortality also  
 153 increases, thus at application dose rate of 1500 mg/kg  $87.3 \pm 3.0$  mortality was achieved.  
 154 Moreover, as the raw DE combined with additional 5mg/ kg permethrin, after seven (7) days  
 155 of exposure the mortality at the lowest dose rate of 500 mg/kg was  $78.9 \pm 4.1$ . As the dose rate  
 156 increases mortality rate also increased; at 1500 mg/ kg of raw DE and 5 mg/kg permethrin  
 157 100% mortality was observed (Table 1)

158  
 159 Table 1: Effect of raw and enhanced DE on mortality of *R. dominica* and adults after 7 days  
 160 of exposure.  
 161

Dose rate (mg/kg)	Mortality (SE±)
0.0	$2.0 \pm 0.6^b$
500	$65.7 \pm 4.7^a$
750	$69.6 \pm 4.3^a$
1000	$75.7 \pm 4.0^a$
1500	$81.1 \pm 3.1^a$
F	78.7
P	< 0.0001
0.0	$1.1 \pm 0.6^c$
500+2mg(Perm)	$66.7 \pm 3.9^b$
750+2mg(Perm)	$71.1 \pm 1.0^b$
1000+2mg(Perm)	$80.1 \pm 0.9^{ab}$
1500+2mg(Perm)	$87.3 \pm 3.0^a$
2mg (permethrin)	$93.8 \pm 6.2^a$
F	105
P	<0.0001
0.0	$3.3 \pm 0.0^c$
500+5mg(Perm)	$78.9 \pm 4.1^b$
750+5mg(Perm)	$87.8 \pm 3.3^{ab}$
1000+5mg(Perm)	$90.0 \pm 5.8^{ab}$
1500+5mg(Perm)	$100.0 \pm 0.0^a$
5mg (permethrin)	$96.7 \pm 3.3^a$
F	110
P	<0.0001

162 Mean within columns and within treatment group followed by the same letter(s) are not  
 163 significantly different ( $p > 0.05$ ) from each other: Tukey-kramer HSD test.

164 **4.2 Effect of raw and enhanced DE on mortality of *R. dominica* adults after 14 days of**  
 165 **exposure.**

166 *R. dominica* mortality after 14 days exposure showed significant differences among  
 167 the different doses of the raw DE and enhanced DE on wheat. After 14 days of exposure  
 168 interval to raw DE, high mortality rate of >80% was observed at the lowest dose rate of 500  
 169 mg/kg. At the highest dose rate of 1500 mg/kg of raw DE 97.8±8% mortality was recorded,  
 170 combination of 2 mg/kg permethrin, mortality rate at the least dose rate of 500 mg/kg was  
 171 increased to 91.1±4.5. Increase in dose rate to 1500 mg/kg complete adult mortality was  
 172 obtained against the test insects. Furthermore, addition of 5 mg/kg permethrin to raw DE,  
 173 further increases the adult mortality to 94.7±5.3. At 1500 mg/kg and 5 mg/kg permethrin,  
 174 complete adult mortality was recorded. However, there was no significant differences in  
 175 mortality level when 2 and 5 mg/kg permethrin was used alone or added to 1000 and 1500  
 176 mg/kg of raw DE (Table 2).  
 177

178 Table 2: Mean Mortality (%±SE) of *R. dominica* adults exposed for 14 days on wheat treated  
 179 with raw and enhanced DE.

Dose rate (mg/kg)	Mortality (SE±)
0.0	3.3±0.2 <sup>b</sup>
500	90.0±5.8 <sup>a</sup>
750	93.3±1.2 <sup>a</sup>
1000	94.4±5.6 <sup>a</sup>
1500	97.8±2.2 <sup>a</sup>
F	116
P	<0.0001
0.0	2.2±0.6 <sup>b</sup>
500+2mg(Perm)	91.1±4.5 <sup>a</sup>
750+2mg(Perm)	96.7±3.3 <sup>a</sup>
1000+2mg(Perm)	97.8±2.2 <sup>a</sup>
1500+2mg(Perm)	100.0±0.0 <sup>a</sup>
2mg (permethrin)	100.0±0.0 <sup>a</sup>
F	245
P	<0.0001
0.0	4.0±1.7 <sup>b</sup>
500+5mg(Perm)	94.7±5.3 <sup>a</sup>
750+5mg(Perm)	96.7±3.3 <sup>a</sup>
1000+5mg(Perm)	100.0±0.0 <sup>a</sup>
1500+5mg(Perm)	100.0±0.0 <sup>a</sup>
5mg (permethrin)	100.0±0.0 <sup>a</sup>
F	210
P	<0.0001

180 Mean within columns and within treatment group followed by the same letter(s) are not  
 181 significantly different (p>0.05) from each other: Tukey-kramer HSD test.

182 **4.3 Effect of raw and enhanced DE on progeny production and percent progeny**  
 183 **inhibition of *R. dominica* adults on treated wheat.**

184 The effect of raw and enhanced DE on progeny production, percentage of dead adult  
 185 progeny and percentage progeny inhibition of *R. dominica* was shown in Table 3. The results  
 186 reveals that there were significant differences (p<0.05) among all dose rates and storage  
 187 periods on numbers of progenies production and inhibition rate of *R. dominica*. After 40 days  
 188 storage period there were only very few 2.5% F<sub>1</sub> adult progeny were emerged, when

189 compared with untreated control 113.7%. On raw DE alone treated grain the percentage of  
 190 dead adult progeny ranged from 69.3±0.5% to 77.6±1.4%, while progeny inhibition ranged  
 191 from 94.1% to 98.2%. After 80 days of storage period only 2.0% adult progeny emerged  
 192 were as in the untreated control with 202.7±4.4 adults developed. At the highest dose rate  
 193 1500 mg/kg only 0.4±0.4% adult progeny were recorded. At all dose rates progeny inhibition  
 194 was >98% compared to the untreated control. Similarly, combination of raw DE with 2  
 195 mg/kg permethrin only 2.3% adult progeny were emerged when compared with untreated  
 196 control being 93.7±6.4% and inhibited 93.6% of F<sub>1</sub> adult progeny at least dose rate of 500  
 197 mg/kg, at 1500mg/ kg only 1.2% adult were developed and inhibited 98.9%. Also after 80  
 198 days of storage, at the lowest application dose rate only 1.9 adults were recorded when  
 199 compared to 192.7±7.1 in the untreated control while F<sub>1</sub> adult progeny inhibition was 98.2%.  
 200 At 1500 mg/kg no adult progenies was observed after 80 days of storage. Moreso, after  
 201 addition of 5mg/kg permethrin, the mean number of F<sub>1</sub> progeny emerged at the least dose rate  
 202 of 500 mg/kg was only 1.9% and adult progeny inhibition rate was 98.4%, at 1500 mg/kg  
 203 DE combined with permethrin dose no F<sub>1</sub> adult progeny were recorded. After 80 days of  
 204 storage period at the least dose rate of 500 mg/kg only very few 2.0% adult progeny were  
 205 emerged whereas 202.7±4.4 F<sub>2</sub> were emerged in the untreated control. Also, at the highest  
 206 dose rate of 1500mg/ kg combined with 2 and 5 mg/kg permethrin and 2 and 5mg/kg alone of  
 207 permethrin dose no adult progenies were developed (Table, 3).

208 **Table: 3 Mean numbers (%±SE) of progeny, percent dead progeny and percent**  
 209 **progeny inhibition of *R. dominica* adults exposed to wheat treated with raw and**  
 210 **enhanced DE.**

40 Days		80 Days					
Dose (Mg/kg)	rate	Mean no. of progeny	% Dead progeny	% progeny inhibition	Mean no. of progeny	% Dead progeny	% progeny inhibition
0.0		113.7±6.4a	2.5±0.4b	-	202.7±4.4a	1.9±0.5c	-
500		2.5±0.1b	69.3±0.5	94.1	2.0±0.3b	75.2±6.3b	98.2
750		2.1±0.2b	71.9±1.6a	95.9	1.6±0.2b	83.0±3.5ab	99.0
1000		1.8±0.2b	75.4±5.2a	97.1	1.5±0.1b	88.8±1.7ab	99.2
1500		1.4±0.0c	77.6±1.4a	98.2	0.4±0.4c	95.8±4.2c	99.9
F		308	160	-	2054	100	-
P		<0.0001	<0.0001	-	<0.0001	<0.0001	-
0.0		93.7±6.0a	3.2±0.8d	-	192.7±7.1a	2.1±0.3d	-
500+2mg(perm)		2.3±0.1b	66.7±0.0c	93.6	1.9±0.0b	81.4±1.4c	98.4
750+2mg(perm)		2.1±0.1b	70.9±2.1bc	95.7	1.6±0.2b	84.3±2.8c	98.9
1000+2mg(perm)		1.8±0.1b	75.0±2.9a	97.1	1.5±0.1b	88.3±1.7bc	99.1
1500+2mg(perm)		1.2±0.0b	95.2±0.1a	98.9	-	96.3±3.7ab	-
2mg(permethrin)		0.0±0.0b	-	-	-	-	-
F		350	537	-	720	298	-
P		<0.0001	<0.0001	-	<0.0001	<0.0001	-
0.0		95.0±7.0a	2.7±0.4	-	213.7±27.4a	3.3±0.2b	-
500+5mg(perm.)		2.0±0.2b	73.5±5.6	96.1	1.8±0.1b	83.0±1.9a	96.1
750+5mg(perm.)		1.9±0.0b	75.0±2.1	96.5	1.2±0.0b	89.3±3.0a	96.5
1000+5mg(perm.)		1.8±0.1b	80.1±0.9	97.2	-	-	-
1500+5mg(perm)		0.0±0.0b	-	-	-	-	-
5mg (permethrin)		0.0±0.0b	-	-	-	-	-
F		175	279	-	60.2	318	-
P		<0.0001	<0.0001	-	<0.0001	<0.0001	-

211 Mean within columns and within treatment group followed by the same letter(s) are not  
 212 significantly different ( $p>0.05$ ) from each other: Tukey-kramer HSD test.

213 **4.4 Effect of raw and enhanced DE on grain damage kernel caused by *R. dominica***

214 The results obtained shows that the percentage of insect damage kernel (IDK) was  
 215 significantly affected by both raw and enhanced DE treatments. There were significant  
 216 differences in the number of damage kernel among different treatment dose rates. Untreated  
 217 control after 80 days of storage complete kernel damage was observed and this was  
 218 significantly higher than values in all other treatments. The result revealed that at the lowest  
 219 dose rate of 500 mg/kg of raw DE 7.0% kernel damage were recorded; and as the dose rate  
 220 increases the rate of damage kernel decreases, in this way very few 3.0% damage kernel were  
 221 observed at 1500 mg/kg. Furthermore, when raw DE combined with 2 mg/kg of permethrin  
 222 dose and applied with 500 mg/kg only 4.0% damaged kernels was recorded. As the dose rate  
 223 increases to 1500 mg/kg only 0.3% kernels were damaged, as against the untreated control  
 224 with complete damaged kernel were noted. More so, combination of 5mg/ kg permethrin dust  
 225 with 500 mg/kg DE only 2.3% kernel were damage. Finally, at the highest dose rate of  
 226 1500mg/ kg no damaged kernel was observed, though there was significant different from  
 227 combined treatments containing 5 mg/kg permethrin with 750 and 1000 mg/kg raw DE with  
 228 1.7 and 0.7% damaged kernels respectively (Table, 4).

229 **Effect of raw and enhanced DE on grain damage caused by *R. dominica*.**

230 Table 4: Mean percent ( $\% \pm SE$ ) of damage kernel caused by *R. dominica* in wheat treated  
 231 with to raw and enhanced DE

Dose rate (mg/kg)	Insect Damage Kernel (IDK) (SE $\pm$ )
0.0	100.0 $\pm$ 0.0 <sup>a</sup>
500	7.0 $\pm$ 0.6 <sup>b</sup>
750	5.3 $\pm$ 0.7 <sup>bc</sup>
1000	4.3 $\pm$ 0.9 <sup>bc</sup>
1500	3.0 $\pm$ 0.7 <sup>c</sup>
F	4847
P	<0.0001
0.0	100 $\pm$ 0.0 <sup>a</sup>
500+2mg(Perm)	4.0 $\pm$ 0.6 <sup>b</sup>
750+2mg(Perm)	2.0 $\pm$ 0.6 <sup>bc</sup>
1000+2mg(Perm)	1.0 $\pm$ 0.6 <sup>c</sup>
1500+2mg(Perm)	0.3 $\pm$ 0.3 <sup>c</sup>
2mg (permethrin)	1.0 $\pm$ 0.6 <sup>c</sup>
F	6701
P	<0.0001
0.0	100.0 $\pm$ 0.0 <sup>a</sup>
500+5mg(Perm)	2.3 $\pm$ 0.3 <sup>b</sup>
750+5mg(Perm)	1.7 $\pm$ 0.2 <sup>bc</sup>
1000+5mg(Perm)	0.7 $\pm$ 0.4 <sup>cd</sup>
1500+5mg(Perm)	0.0 $\pm$ 0.0 <sup>d</sup>
5mg (permethrin)	0.0 $\pm$ 0.0 <sup>d</sup>
F	35630
P	<0.0001

232 Mean within columns and within treatment group followed by the same letter(s) are not  
 233 significantly different ( $p>0.05$ ) from each other: Tukey-kramer HSD test.

234 **5.0 Discussion**

235 The result of the present study has showed that raw DE and raw DE enhanced with  
236 permethrin could be used against *R. dominica* on stored wheat. Both raw DE alone and raw  
237 DE combined with permethrin dust caused high adult mortality in this insect specie. Adult  
238 mortality increased with increase in dose rate and exposure period. The results are in  
239 agreement with the finding of Athanassiou *et al.* (2005) and Wakil *et al.* (2010) who reported  
240 that mortality increased with increase in dose rate and exposure period. An increase in  
241 exposure time period to commercial inert DE formulations was shown to increase mortality  
242 of stored product beetles (Arthur, 2000; Subramanyan and Roesli, 2000; Athanassiou *et al.*,  
243 2003). In addition to dessication, it has been reported that DE also reduces the locomotion  
244 ability of stored product insects (Vardeman *et al.*, 2007). Longer exposure interval or DE  
245 higher dose rate needed to achieve 100% mortality for adult of the insect tested (Fields and  
246 Korunic, 2000; Athanassiou *et al.*, 2004).

247 The raw DE caused 95-100% adults mortality after 14 days of exposure. The results  
248 agrees with Kabir *et al.*(2010, 2011) who reported that raw DE was effective against *R.*  
249 *dominica* (F.) at higher dose rates 1500 mg/kg or above. Similarly, Mvumi *et al.* (2008)  
250 found that African DE applied at higher dose rate shows significantly high mortality of  
251 *Sitophilus zeamais*, *T. castaneum* and *R. dominica* after 7, 14 and 28 days exposure period.  
252 Hence the result of this study suggest that relatively higher dose 1500 mg/kg or above is  
253 required to suppress *R. dominica* and after 14 days in stored wheat. This rate was more than  
254 the labeled rate for commercial DE products. For instance, the labeled rate for Protect It<sup>®</sup>,  
255 Insecto<sup>®</sup> and Dryacide are 400, 500 and 1000 ppm respectively (Vardeman *et al.*, 2007). In  
256 another earlier study, Mvumi *et al.* (2006) noted that African DE samples were unsuitable as  
257 grain protectant at dose rate of 1000 ppm. The explanation was that samples contain high  
258 level of contaminants and were not collected from deep enough layers of DE deposits. In the  
259 case of the raw DE tested in this study, the researcher was not involved in the sample  
260 collection of DE. Nevertheless, high efficacy was observed against *R. dominica* which  
261 sustained 97.8 % adult mortality. Progeny suppression at dose rate 1500 mg/kg on wheat was  
262 almost 100%. Thus, the present DE seems to be more effective against stored grain insects  
263 than those studied by Mvumi *et al.* (2006).

264 The general trend of efficacy observed in the present study was similar to those  
265 reported by other authors. According to Athanassiou and Korunic (2007) progeny production  
266 in the treated commodity is perhaps more important than parental mortality because a grain  
267 protectant should protect the grain for a long storage period. The most important finding of  
268 this study is that the raw DE product used affect the test insect species in same way as  
269 commercial DE formulations.

270 The number of progeny developed and their subsequent survival was significantly  
271 affected by DE dose alone and in combination with permethrin dust and storage period.  
272 Noticeable reduction in progeny production was obtained with increase in dose rates and  
273 storage period. Furthermore, complete progeny inhibition was not observed at 40 days of  
274 storage in all the treatments. In a test, using Protect It<sup>®</sup> against *S. oryzae*, Arthur and Throne  
275 (2003) reported that progenies emerged in treatment where 100% mortality of exposed adult  
276 was observed, and this could be attributed to the mode of action of DEs. It appears that stored  
277 product insect can lay eggs following initial exposure to DE before subsequent lethality  
278 manifest.

279 **6.0 Conclusion**

280 This present study document that raw DE alone and in combination with 2 and 5  
281 mg/kg permethrin has insecticidal potential in control of *R. dominica*. The efficacies varies  
282 with dose rate and exposure periods. However, additions of permethrin increases potency of  
283 raw DE. *R. dominica* could be effectively control with raw DE at dose rate of 1500 mg/kg  
284 raw DE at 1000 mg /kg + 5 mg permethrin or 1500 mg + 2 mg permethrin may required.  
285 Complete prevention of grain damage by *R. dominica* could be achieved at 1500 mg raw  
286 DE+ 5mg permethrin.

## 287 **7.0 Recommendations**

288 Based on the findings of this research it is possible to recommend 1500 mg/kg of raw  
289 DE combined with 5 mg/kg permethrin dust for the control of *R. dominica* in stored wheat.  
290 Further investigations are recommended to investigate potential effective dosage in  
291 controlling other stored product insects. The use of DE in management of stored product  
292 insects in Nigeria should be encourage as their use is safe both humanly and environmentally.

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